11330/1 NREAD 18 Oct 84

From: Director, Natural Resources and Environmental Affairs Division, Marine Corps Base, Camp Lejeune To: Base Maintenance Division, Marine Corps Base, Camp Lejeune

Subj: DRINKING WATER ANALYSIS; CERTIFICATION FOR

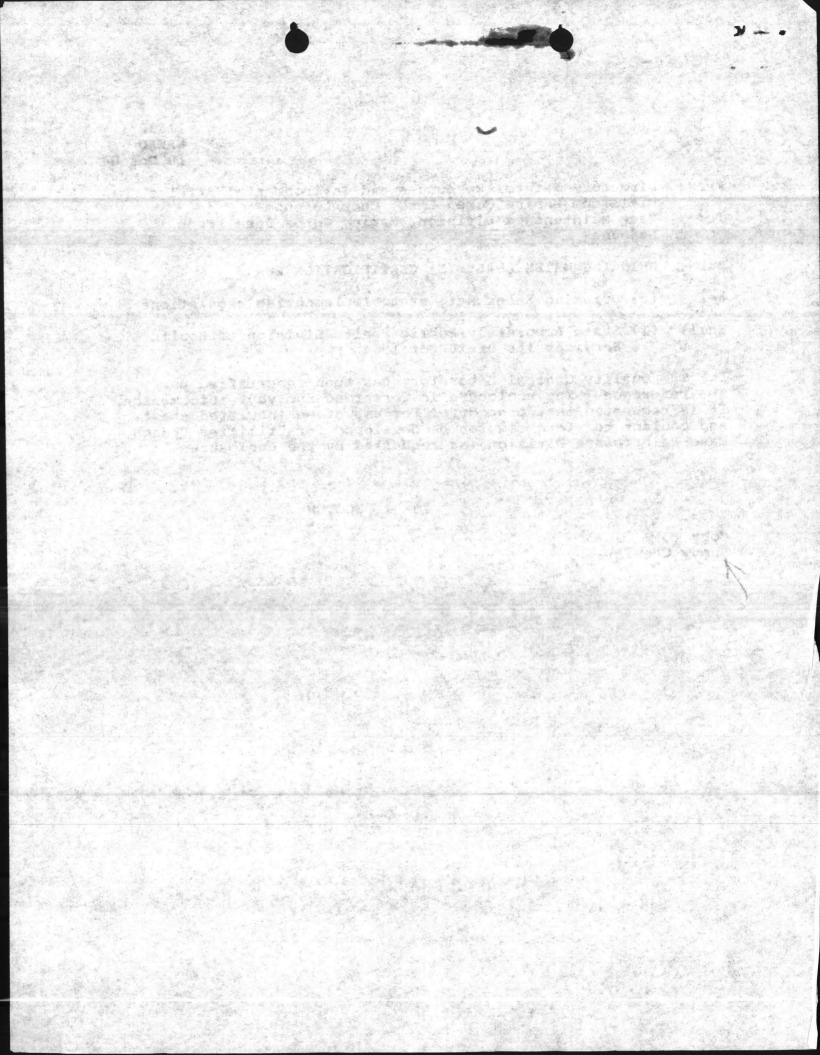
Ref: (a) Drinking Water Act; state implementing regulations

Encl: (1) State Laboratory Public Bealth Division of Health Services 1tr of 10 Oct 1984

1. The Quality Control Laboratory has been recertified per the reference. The enclosure is forwarded for your information. It is requested that procedures for use of an insulated chest and coolant to store samples be developed for Utilities Branch, Base Maintenance Division, as requested by the enclosure.

J. I. WOOTEN

Copy to: Supvy Chemist



Ronald H. Levine, M.D., M.P.H. STATE HEALTH DIRECTOR

DIVISION OF HEALTH SERVICES STATE LABORATORY OF PUBLIC HEALTH 306 N. Wilmington St. P.O. Box 28047 Raleigh, N.C. 27611-8047

October 10, 1984

Commanding General Marine Corps Base Camp LeJeune, NC 28542

ATTN: Facilities - NREAD Elizabeth Betz

Dear Ms. Betz:

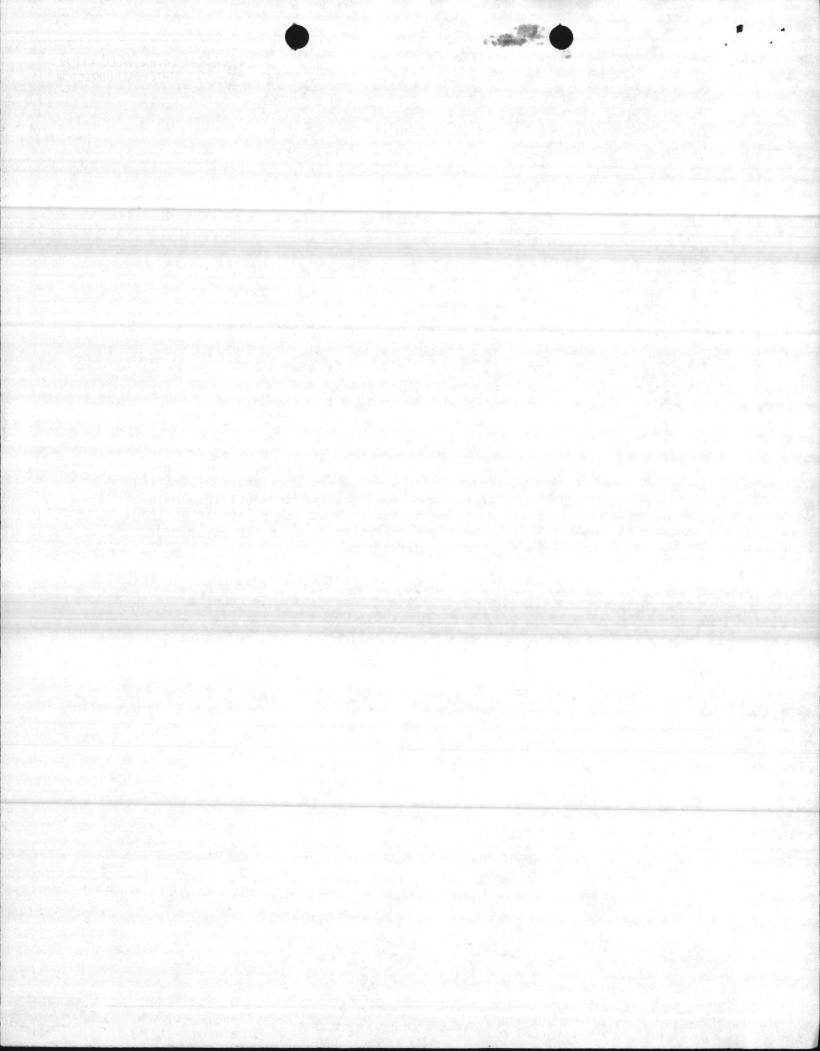
The findings of the on-site evaluation on Sept. 27, 1984 indicate that your laboratory has met the minimum requirements for certification as specified in North Carolina Drinking Water Regulations (IONCAC 9D .0301 - .0330). We, therefore, grant Certification to your laboratory for total coliform analysis on public water supplies.

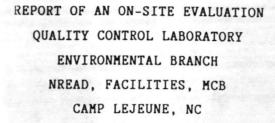
If you have any questions or if we may be of further assistance in this matter, please let us know.

Sincerely,

E. D. Beesley Laboratory Certification Evaluator

EDB/hw Enclosure

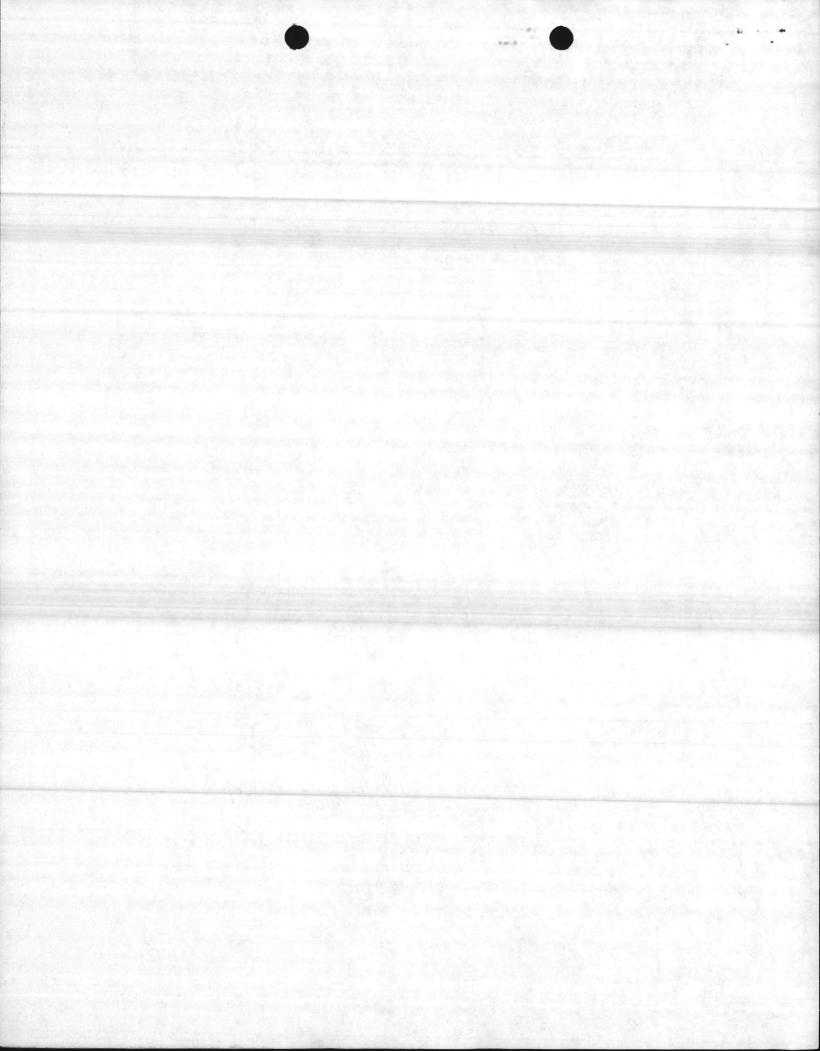


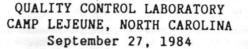


SEPTEMBER 27, 1984

BY: E. D. BEESLEY LABORATORY CERTIFICATION EVALUATOR ENVIRONMENTAL SCIENCES BRANCH

LABORATORY SECTION NORTH CAROLINA DIVISION OF HEALTH SERVICES NORTH WILMINGTON STREET RALEIGH, NORTH CAROLINA 27611





I. INTRODUCTION

The equipment and procedures employed in the bacteriological analyses of water by this laboratory conformed with the provisions of the North Carolina Safe Drinking Water Regulations, except for the items indicated.

II. DEVIATIONS AND RECOMMENDATIONS

No Deviations.

<u>Recommendation</u>: Samples must be protected from excessive heat, especially during hot summer months. An insulated chest containing a coolant such as frozen Blue Ice should be used.

III. LIST OF PERSONNEL

NAME

POSITION

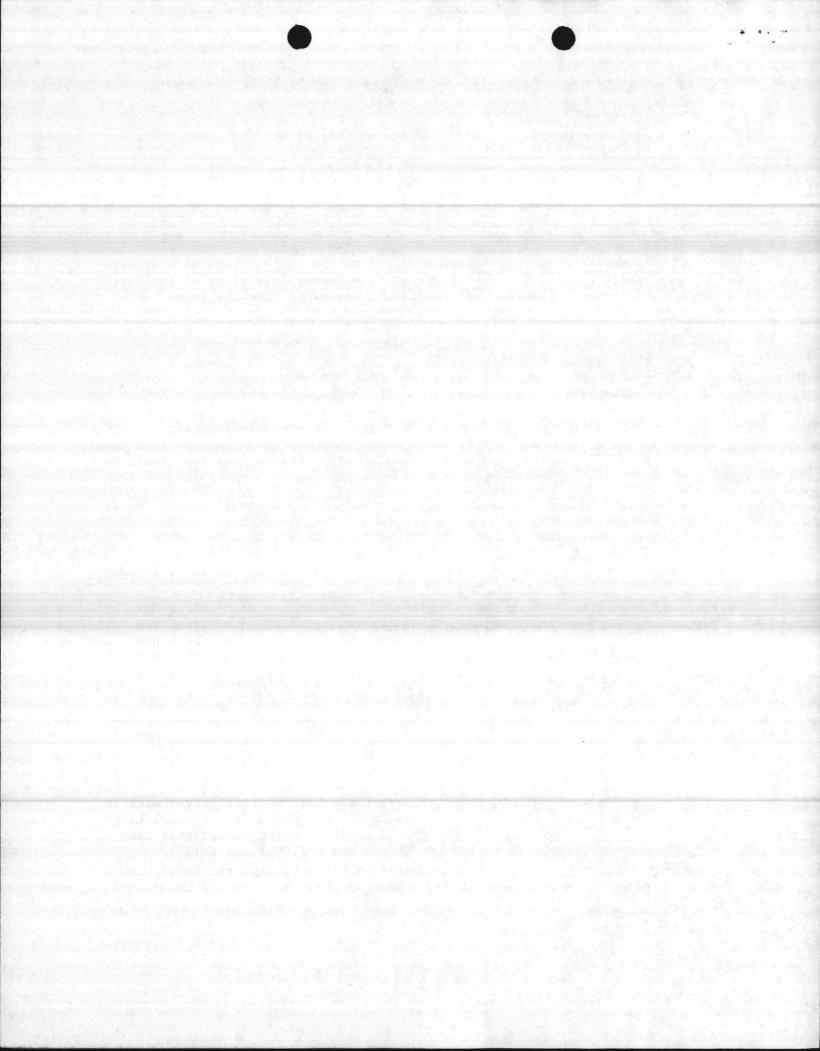
Elizabeth A. Betz Hoy Burns Bob Lachapelle Gaines Honeycutt Thomas Barbee

Supervisor Techn./Analyst Techn./Analyst Techn./Analyst Techn./Analyst TEST NORMALLY PERFORMED

MF & MPN MF & MPN MF & MPN MF & MPN MF & MPN

V. CONCLUSION:

The procedures and equipment in use at the time of this survey were in general compliance with the provisions of the North Carolina Drinking Water Regulations (10NCAC 9D .0301 - .0330). We recommend that the analytical data be accepted for MF and MPN Coliform Analysis of Drinking Water under the North Carolina Safe Drinking Water Act.



North Carolina Department of Human Resources Division of Health Services



Certification for the analysis of drinking water has been granted to

CAMP LEJEUNE QUALITY CONTROL LABORATORY

for the following parameters

Coliform Bacteria - by Membrane Filter Procedure Coliform Bacteria - by Most Probable Number Procedure

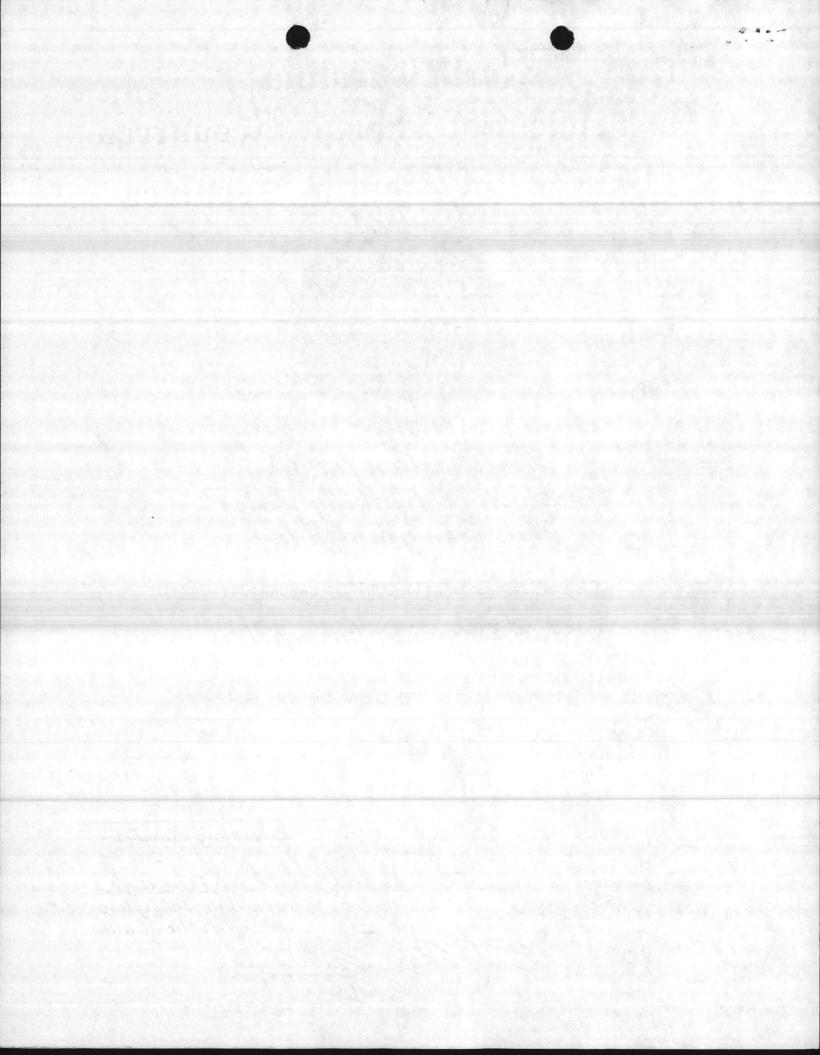
September 1986

Expiration Date Realth Director

37807 Laboratory Number

State Laborflory of Public Health

Certification O



Ronald H. Levine, M.D., M.P.H. STATE HEALTH DIRECTOR

DIVISION OF HEALTH SERVICES STATE LABORATORY OF PUBLIC HEALTH 306 N. Wilmington St. P.O. Box 28047 Raleigh, N.C. 27611-8047

October 10, 1984

Commanding General Marine Corps Base Camp LeJeune, NC 28542

ATTN: Facilities - NREAD Elizabeth Betz

Dear Ms. Betz:

The findings of the on-site evaluation on Sept. 27, 1984 indicate that your laboratory has met the minimum requirements for certification as specified in North Carolina Drinking Water Regulations (10NCAC 9D .0301 - .0330). We, therefore, grant Certification to your laboratory for total coliform analysis on public water supplies.

If you have any questions or if we may be of further assistance in this matter, please let us know.

Sincerely,

E. D. Beesley Laboratory Certification Evaluator

EDB/hw Enclosure

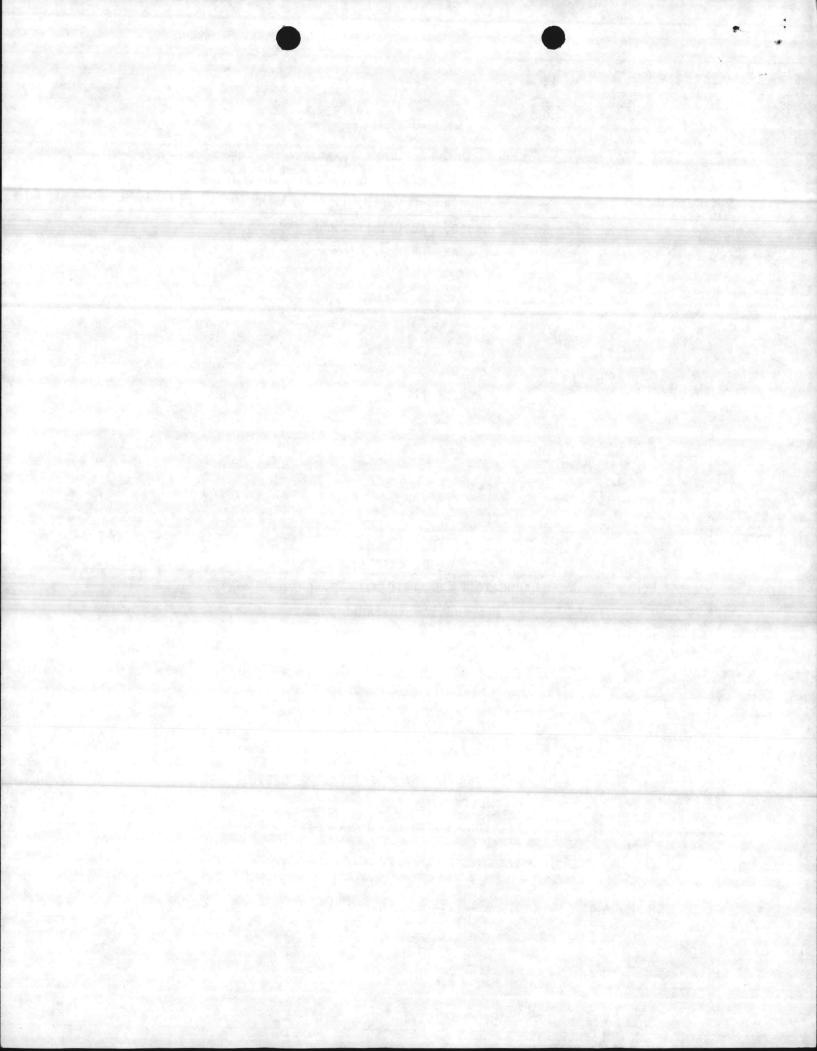
REPORT OF AN ON-SITE EVALUATION QUALITY CONTROL LABORATORY ENVIRONMENTAL BRANCH NREAD, FACILITIES, MCB CAMP LEJEUNE, NC

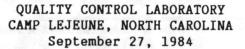
SEPTEMBER 27, 1984

BY:

E. D. BEESLEY LABORATORY CERTIFICATION EVALUATOR ENVIRONMENTAL SCIENCES BRANCH

LABORATORY SECTION NORTH CAROLINA DIVISION OF HEALTH SERVICES NORTH WILMINGTON STREET RALEIGH, NORTH CAROLINA 27611





I. INTRODUCTION

The equipment and procedures employed in the bacteriological analyses of water by this laboratory conformed with the provisions of the North Carolina Safe Drinking Water Regulations, except for the items indicated.

II. DEVIATIONS AND RECOMMENDATIONS

No Deviations.

<u>Recommendation</u>: Samples must be protected from excessive heat, especially during hot summer months. An insulated chest containing a coolant such as frozen Blue Ice should be used.

III. LIST OF PERSONNEL

NAME

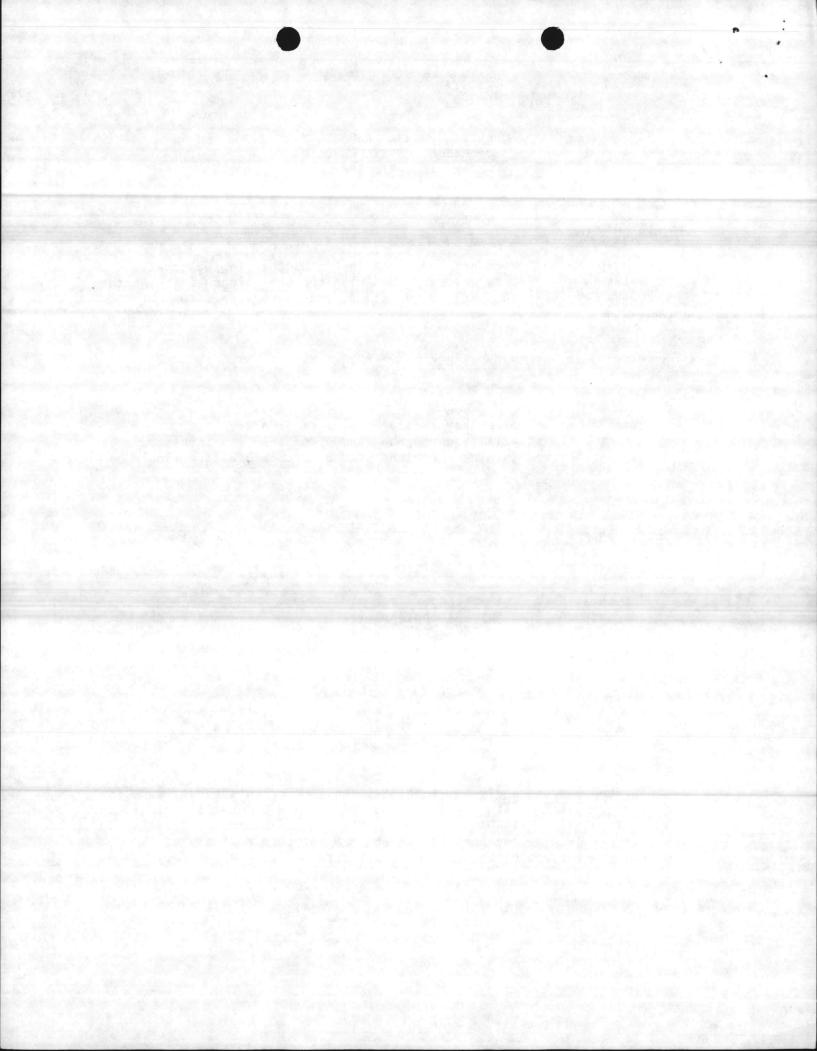
POSITION

TEST NORMALLY PERFORMED

Elizabeth A. Betz	Supervisor	MF & MPN
Hoy Burnsdford	Techn./Analyst	MF & MPN
Bob Lachapelle	Techn./Analyst	MF & MPN
Gaines Honeycutt	Techn./Analyst	MF & MPN
Thomas Barbee	Techn./Analyst	MF & MPN

V. CONCLUSION:

The procedures and equipment in use at the time of this survey were in general compliance with the provisions of the North Carolina Drinking Water Regulations (10NCAC 9D .0301 - .0330). We recommend that the analytical data be accepted for MF and MPN Coliform Analysis of Drinking Water under the North Carolina Safe Drinking Water Act.



STATE LABORATORY OF PUBLIC HEALTH

DIVISION OF HEALTH SERVICES

NORTH CAROLINA DEPARTMENT OF HUMAN RESOURCES

P. O. BOX 28047, 306 NORTH WILMINGTON STREET,

RALEIGH, NORTH CAROLINA 27611

FORMS FOR ON-SITE EVALUATION OF LABORATORIES INVOLVED IN ANALYSIS OF PUBLIC WATER SUPPLIES

MICROBIOLOGY

LAB I.D. NUMBER: 37807

LABORATORY: Camp LeJeune Quality Control Laboratory - Environmental Branch

STREET: NREAD, Facilities, MCB

CITY: Camp LeJeune STATE: NC 28542

TELEPHONE NUMBER: 919-451-5977

SURVEY BY: E. D. Beesley

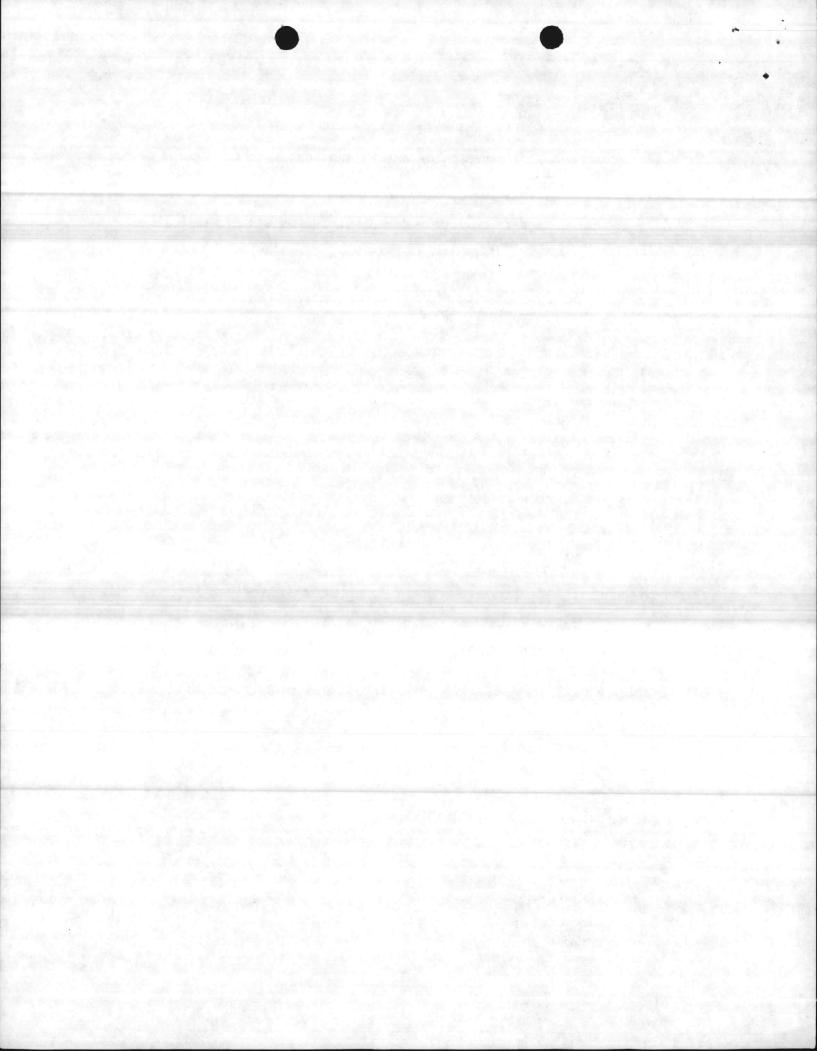
AFFILIATION: North Carolina Division of Health Services

DATE: September 27, 1984

Codes for Marking On-Site Evaluation Forms:

S - Satisfactory X - Unsatisfactory NA - Not Applicable

DHS 3068 Laboratory







PERSONNEL

POSITION/	NAME	I	CADEMI	C TRAIN	NING	TESTING	EXPERIENCE
TITLE		HS	BA/BS	MA/MS	PH.D	METHOD(S)	(YEARS/AREA)
Laboratory Director		Stratt Line					
Supervisor	Elizabeth A. Betz*	x	BS Chem.			MF & MPN	5 years
	Hoy Burns* Bob Lachapelle*	x				MF & MPN MF & MPN	8 years
Technician/ Analyst	Gaines Honeycutt* Thomas Barbee	x x x	BS			MF & MPN MF & MPN	3 years 3 years

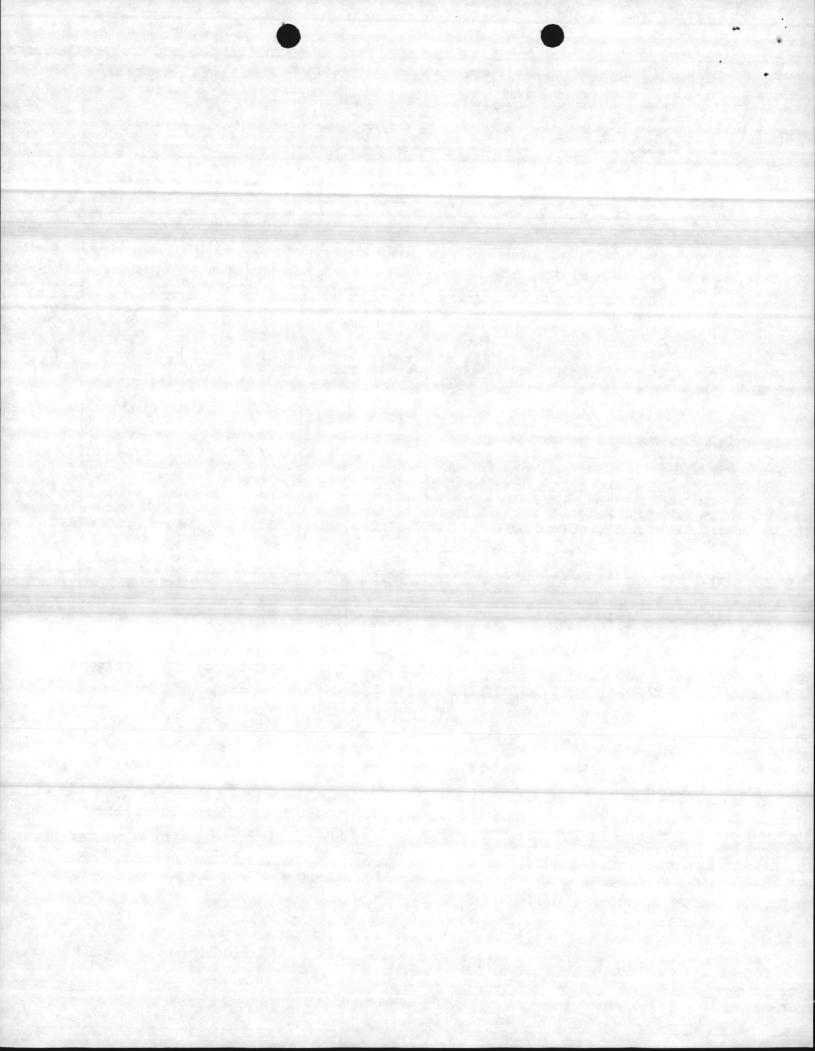
* Attended NCDHS Watermicro Workshop

LABORATORY FACILITIES

Space in laboratory and preparation room is adequate for needs during peak. work periods (200 ft and 6 linear ft. of usable bench space per analyst).

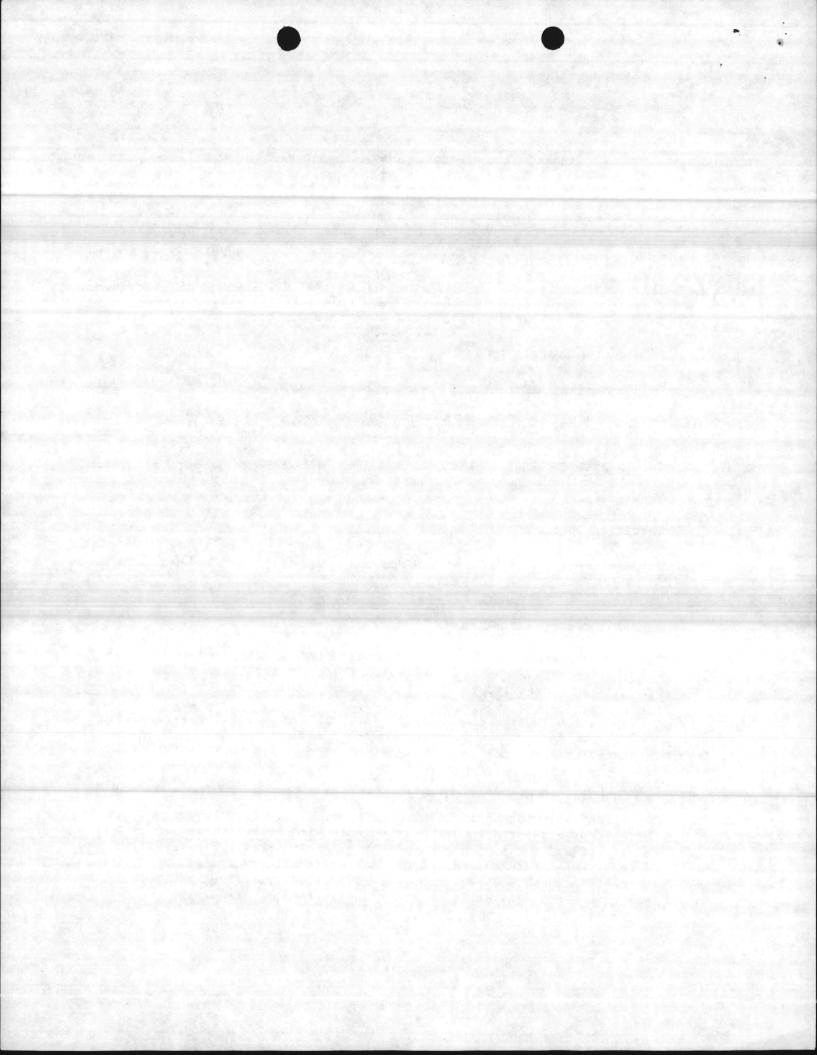
Facilities are clean, with adequate lighting (100 ft-candles) and air-conditioning.

Satisfactory



LABORATORY EQUIPMENT, SUPPLIES, AND MATERIALS

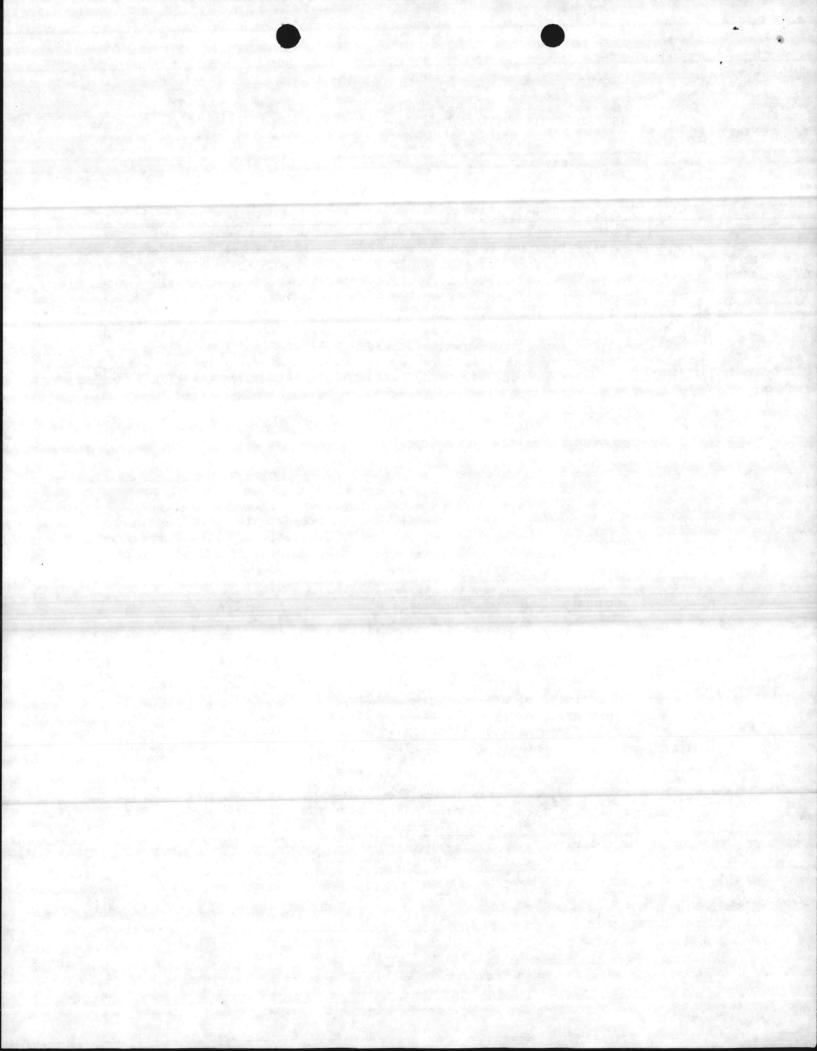
	pH Meter			70	
	Manufacturer	Orion Orion	Mode	1 61	1
	Aliquot of star	ted to 0.1 pH units each ndard pH 7.0 buffer used repared buffer dated when	only once		
	Balance-Top Lo	ader or Pan			
	Manufacturer	Ohaus	Mode	el <u>Harva</u>	ard Trip
	preparatio	s a 50-mg weight accurat n of 2-g quantities) ually eights in clean conditio	· · · · · · · · · · · · · · · · · · ·	 	
•	Thermometers				
	Certified Ther	mometer:			
	Manufacturer	Fisher (Kessler)	Certificate	# 836-2	12
	. Maximum Regist	ering:			
	Manufacturer	Brooklyn			
		ters calibrated annually	against a certifi	ed rmometer	rs
	thermomete checked qu Legible gradua	er or one of equivalent a marterly tions in liquid column		••••••	••••• <u>S</u>
	thermomete checked qu Legible gradua	arterly tions in liquid column		••••••	••••• <u>S</u>
	thermomete checked qu Legible gradua No separation	arterly tions in liquid column			••••• <u>S</u>



LABORATORY EQUIPMENT, SUPPLIES, AND MATERIALS (Continued)

5. Autoclave

	Manufacturer Market Forge Model Sterlimatic
	Reaches sterilization temperature (121°C), maintains 121°C during sterilization cycle, and requires no more than 45 min. for a
	complete cycleS Temperature checked at least weekly with a maximum registering thermometer calibrated in 1°C increments
	and results recordedS Pressure and temperature gauges on exhaust side and an operating safety valveS
	No air bubbles produced in fermentation vials during depressurization. S Record maintained on time and temperature for each sterilization
	cycleS
6.	Hot-Air Oven
	Manufacturer NA Model
	Operates at a minimum of 170°C Thermometer inserted or oven equipped with temperature-recording thermometer device Time and temperature record maintained for each sterilization cycle
	Thermometer bulb in sand (optional)
7.	Refrigerator
	Temperature maintained at 1° to 5.0°CS
8.	Inoculation Equipment
	Sterilized loops of at least 3-mm, diameter, 22 to 24 gauge Nichrome, Chromel, or platinum-iridium wireS Disposable, dry heat-sterilized, hardwood applicator sticks or presterilized loopsNA
9.	Optical Equipment
	Low power magnification device (preferably <u>binocular microscope</u> with 10 to 15X) with <u>fluorescent light</u> source for counting
	MF colonies <u>S</u> Colonies counted with a mechanical hand tally (optional)
10.	Membrane Filtration Equipment
	Manufacturer Millipore Model
	Made of stainless steel, glass, or autoclavable plastic
	Nonleaking and uncorroded



LABORATORY EQUIPMENT, SUPPLIES, AND MATERIALS (Continued)

11. Membrane Filters and Pads

Manufacturer	Millipore	Туре	HAWG
		The second s	

Filters recommended by manufacturer for water analyses......S Filters and pads presterilized or autoclavable.....S Lot numbers and dates of receipt of membrane filters recorded (optional)

12. Glass, Plastic, and Metal Utensils for Media Preparation

Washing process provides glassware free of toxic residue as demonstrated by the inhibitory residue test and results recorded......S

Detergent: Disperse (American Scientific)

Glass items of borosilicate, free of chips and cracks.....<u>S</u> Utensils clean and free from foreign residues or dried medium.....<u>S</u> Plastic items clear with visible graduations.....<u>S</u>

13. Sample Bottles

Wide-mouth hard glass bottles; stoppered or plastic screw-capped; capacity at least 120 ml	. s
Glass-stoppered bottles with tops covered with aluminum foil or	
	. NA
Whirl-Pak Bags	. NA
Screw-caps have leakproof nontoxic liners that can withstand repeated sterilization (30 min at 121°C)	
Sterility of each batch of sample containers checked using non- selective broth and results recorded	

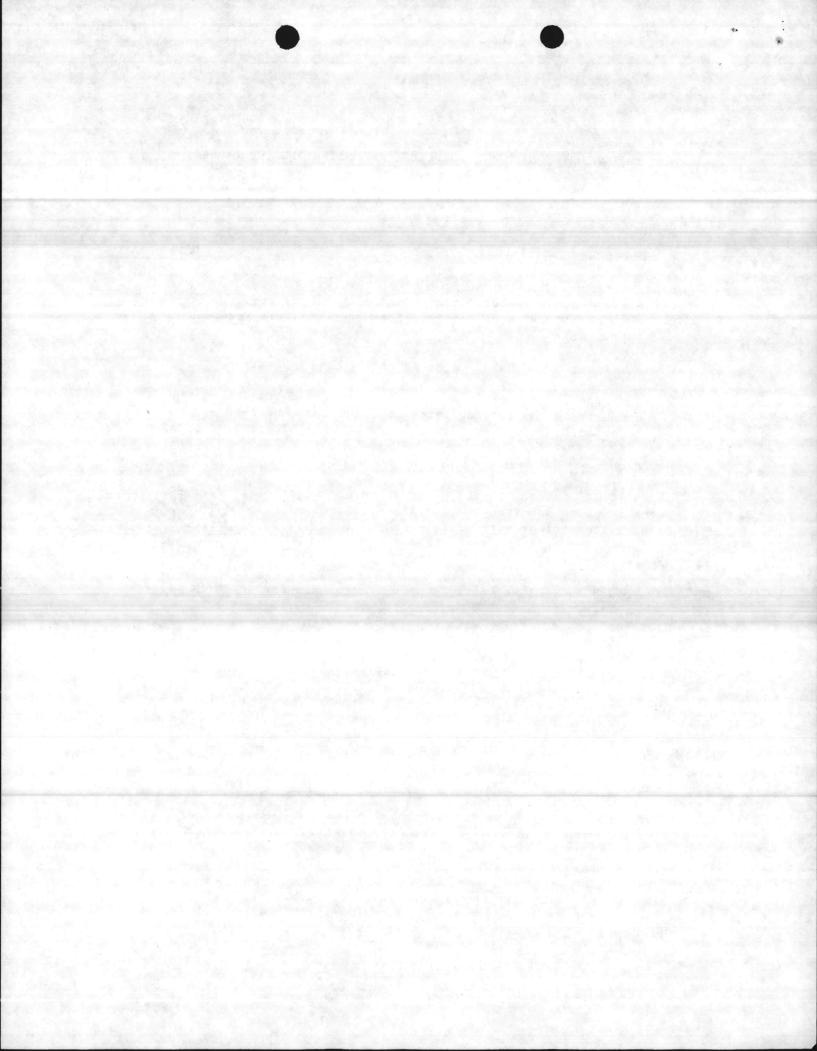
14. Pipets

Brand	Falcon	Туре
	A Present	

Sterile;	glass o	r plastic;	with a 2.	5 percent	tolerance	5
Tips unbr	oken; gr	aduations	distinctly	marked		<u>S</u>

15. Pipet Containers

Aluminum or stainless steel.....<u>NA</u> Pipets wrapped in quality kraft paper (char-resistant).....<u>NA</u> Open packs of disposable sterile pipets resealed between uses.....<u>S</u>



LABORATORY EQUIPMENT, SUPPLIES, AND MATERIALS (Continued)

16. Culture Dishes

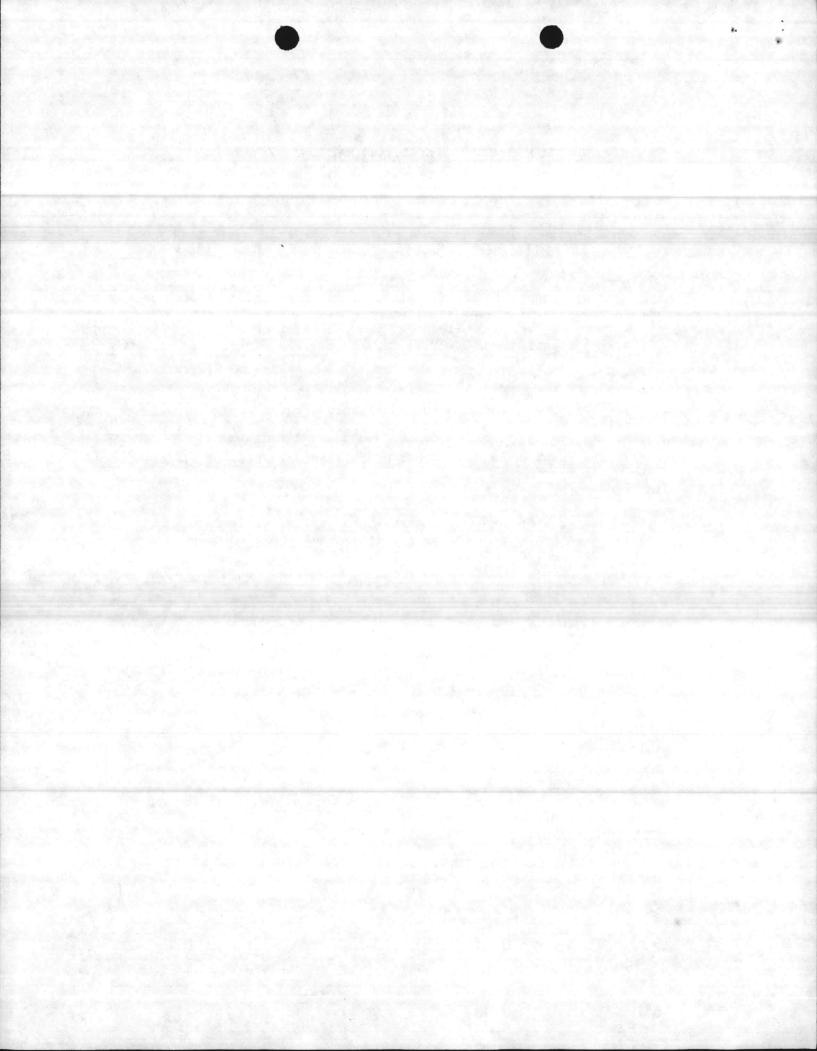
				49 A 9
Brand	Millipore	- Pyrex	Туре	100 X 15
Sterile pla	stic or glass			S
Open packs	of disposable s	terile plastic	dishes resealed	between uses.S
Dishes are	in containers o	f aluminum or	stainless-steel w	ith
covers	or are wrapped	with heavy alu	minum foil or	
char-re	sistant paper			S

17. Culture Tubes and Closures

Sufficient size to contain medium and sample without danger of
spillageS
Metal or plastic capsS
Borosilicate glass or other corrosion-resistant glassS

18. Maintenance

Service conti	racts or approved	internal prot	ocol maintained on	
balance,	autoclave, water	still, etc.;	service records entered	ł
in a log	book			S



GENERAL LABORATORY PRACTICES

1. Sterilization Procedures

Timing for sterilization begins when autoclave reaches 121°C......S Tubed broth media and reagents sterilized at 121°C 12 to 15 min.....S Tubes and flasks packed loosely in baskets or racks for uniform

heating and cooling......S Total exposure of MPN media to heat not over 45 min.....S Dilution water blanks autoclaved at 121°C for 30 min.....S Rinse water volumes of 500 to 1,000 ml sterilized at 121°C for 45 min. MF presterilized or autoclaved at 121°C for 10 min fast exhaust.....S MF assemblies and empty sample bottles sterilized at 121°C

for 30 min	
MF assemblies sterilized between sample filtration series	. <u>s</u>
Wire loops, needles, and forceps sterilized	
Individual glassware items autoclaved at 121°C for 30 min	. s
Individual dry glassware items sterilized 2 hours at 170°C (dry heat)	. <u>s</u>
Pipets, culture dishes, and applicator sticks in boxes sterilized	c

at 170°C for 2 hours..... MPN media removed and cooled as soon as possible after sterilization and stored in cool dark place (optional)

UV light or boiling water for at least 2 min may be used on membrane filter assemblies to reduce bacterial carry-over between each filtration (optional)

Heat-sensitive tapes and/or strips/ampoules used during sterilization (optional)

2. Laboratory Pure Water

Only laboratory pure water, u	sed in pr	ceparing media,	reagents,	rinse	
water, and dilution water					S
Laboratory pure water not in	contact w	with heavy meta	ls		S
Source: Laboratory-prepared	~	Purchas	sed		
If Laboratory-prepared:	Comina	Magapura 61 ar			

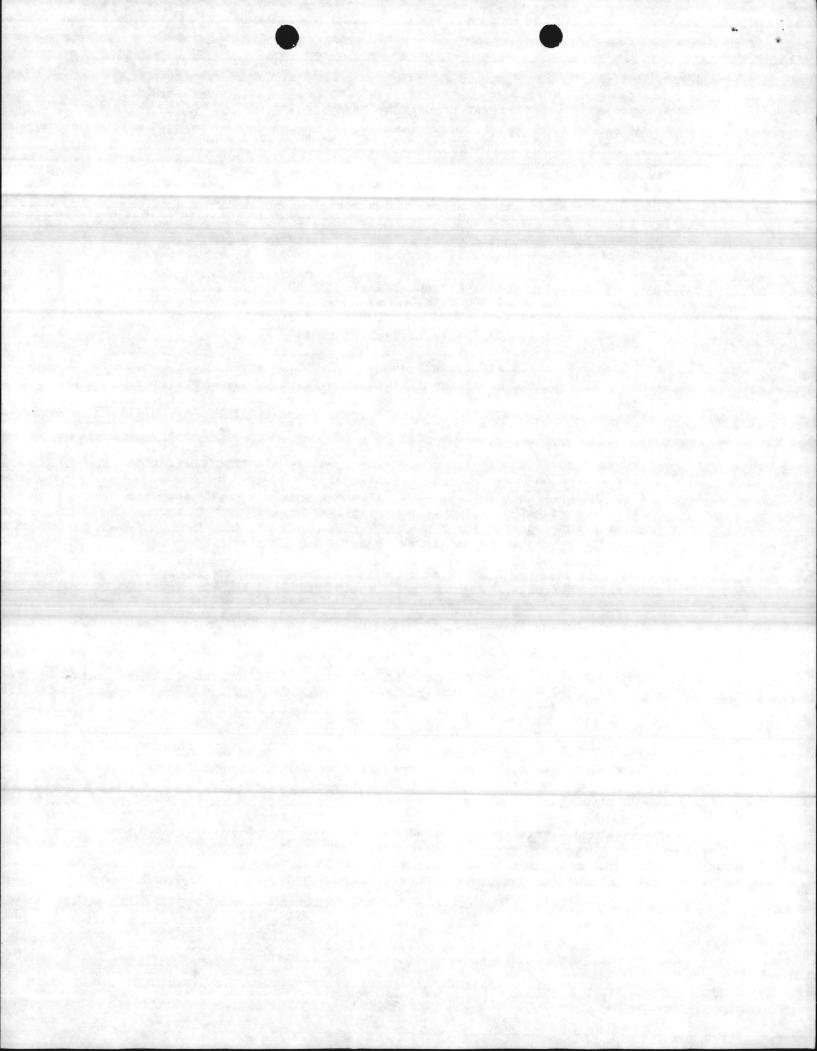
Still Manufacturer	Corning	Megapure	6L and	3L
Deionizer Manufacture	r Cori	ning High	Сар	

Record of recharge frequency

Production rate and quality adequate for laboratory needs......S Inspected, repaired, cleaned by service contract or in-house service...S

a. Chemical Quality Control

Record of satisfactory annual analyses for trace met Chromium, Copper, Nickel, Lead, and Zinc)	tals (Cadmium,
A single metal not greater than 0.05 mg/1 Total metals: equal to or less than 1.0 mg/1	
Testing laboratory GLI	
Record of monthly analyses of laboratory pure water	
Conductance: >0.5 megohm resistivity or <2.0 mic pH: 5.5 - 7.5	S
Standard plate count: <10,000/ml. Stored or de	eionized;
<pre><1000, freshly distilled or ultra-pure Free chlorine residual: <0.1</pre>	



GENERAL LABORATORY PRACTICES (Continued)

b. Microbiological Quality Control

Test for bactericidal properties o	f distilled water	-
(0.8 - 3.0) performed at least	annually	
Testing laboratory	Date	Ratio

3. Rinse and Dilution Water

Stock buffer solution prepared according to "Standard Methods", 14th edition, or EPA Methods Manual.....<u>S</u> Stock buffer solution adjusted to pH 7.2.....<u>S</u> Stock buffer autoclaved at 121°C, stored at 1° to 5.0°C or filter sterilized.....<u>S</u>

Stock buffer labeled and dated.....S Stock potassium phosphate buffer solution (1.25 ml) added per liter distilled water for rinse and dilution water.....S

TILCE GIOC.							CALLER AND	_
Final pH 7.2 +	0.1						<u>S</u>	
MgSO4	MgCl ₂	5	ml	stock	solution	per	liter	_

4. Media

Dehydrated media bottles kept tightly closed and protected from

dust and excessive humidity in storage areasS
Dehydrated media not used if discolored or cakedS
Laboratory pure water used in media preparationS
Media completely dissolved before dispensing to culture
tubes or bottlesS
.MPN tube media with loose-fitting caps used in less than 1 weekS
Tube media in screw-capped tubes held no longer than 3 monthsS
Media stored at low temperatures are incubated overnight prior
to use and tubes with air bubbles discardedS
Media protected from sunlightS
MF media stored in refrigerator; broth media used within 96 hours,
agar within two weeks if prepared in tight-fitting dishesS
Ampouled media stored at 1° to 5.0°C and time limited to
manufacturer's expiration dateS

5. Quality Control of Media and Reagents

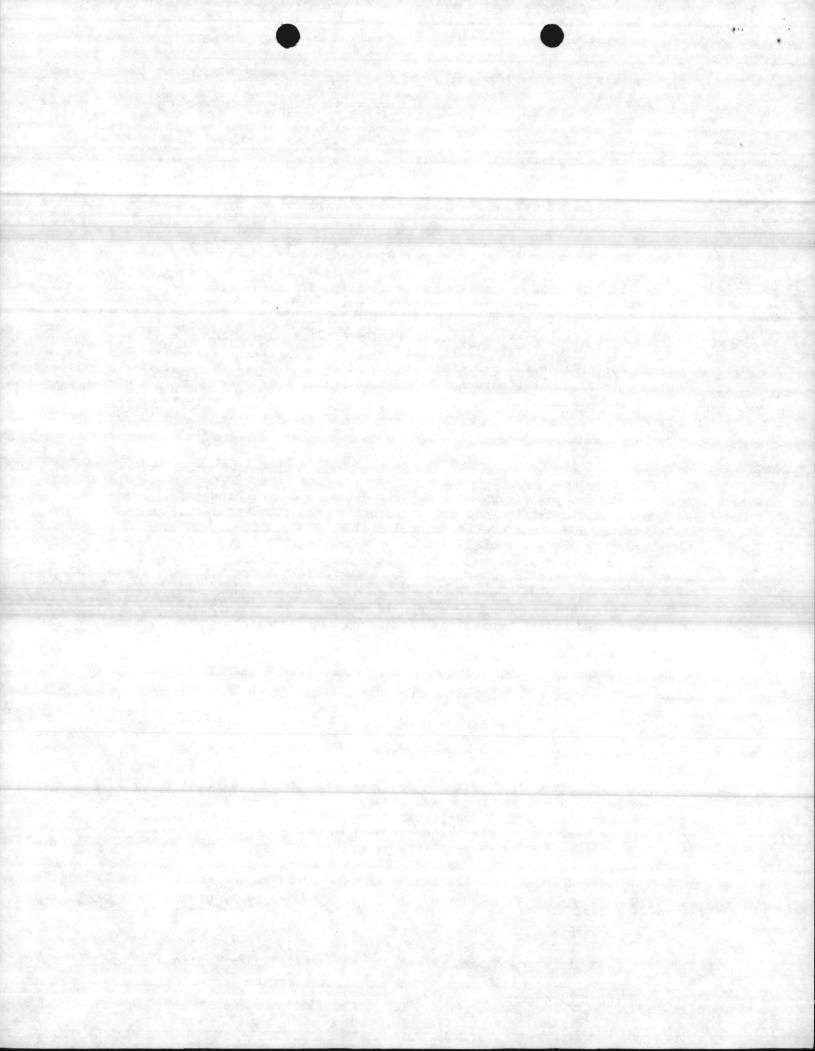
Satisfactory 1	records	containing	complete	quality	control	checks
----------------	---------	------------	----------	---------	---------	--------

on media available for inspection.....<u>S</u> Laboratory chemicals of Analytical Reagent Grade.....<u>S</u> pH checked and recorded on each batch of medium after preparation and after sterilization.....<u>S</u>

Causes for deviations beyond \pm 0.2 pH units specified......S Media ordered on a basis of 12-month need; purchases in 1/4 lb.

quantities, except those used in large amounts (optional) Bottles dated on receipt and when opened (optional) Opened bottles of routinely used media discarded within 6 months (if

stored in desiccator storage may be extended) (optional) Shelf life of unopened bottles not in excess of 2 years (optional) New lots of media quality tested against satisfactory lot using natural water samples (optional)



GENERAL LABORATORY PRACTICES (Continued)

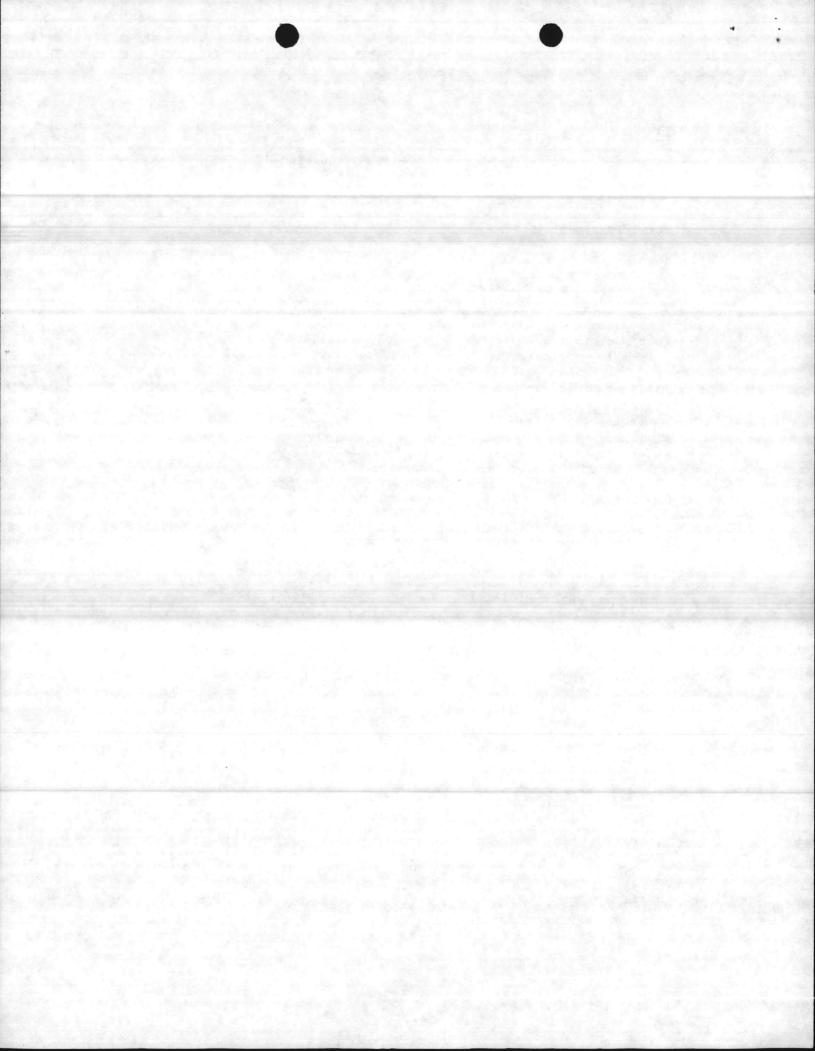
6. Lauryl Tryptose Broth

7.

8.

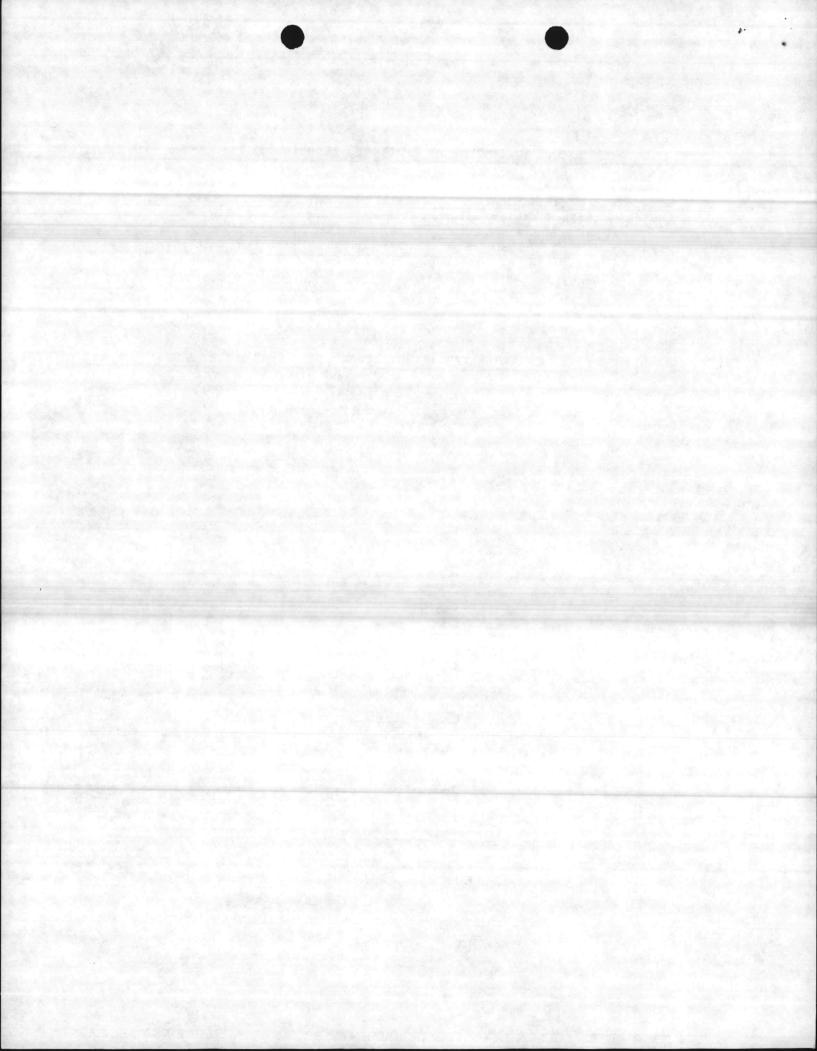
9.

Manufacturer	Difco		Lot No.	715430	2/89
Single strength Not less than 1	pH 6.8 + 0.2; d 0 ml per tube	6.6g per liter pure v louble strength pH 6	$.7 \pm 0.2$.		S
Media made to r	esult in single	strength after addi	tion of		
Brilliant Green	Lactose Bile Br	roth			
Manufacturer	Difco		Lot No.	722173	2/89
Medium composit Final pH 7.2 <u>+</u>	ion 40g per lite 0.2	er pure water	·····		<u>s</u>
M-Endo Media					
Manufacturer	BBL		Lot No.	H6DNQK	8/87
 15g agar ad Reconstituted i	ded/1 n laboratory pu	iter pure water; opt re water containing	2 percent	E	
Final pH 7.2 +	0.2	ath until completely			•••• S
Standard Plate	Count Agar				
Manufacturer	Difco (new)	lot ordered)	Lot No	.788117	
Storilo modium	not remelted a	nd pH 7.0 ± 0.2	erilizat	ion	5
Culture dishes	incubated 48 how 0 ml or less the	urs at $35^{\circ} \pm 0.5^{\circ}$ C an 0.1 ml sample pla	ted (sam	 ple	
or dilution	10 ml or more:	medium temperature	between		
1.10 to 1600	• • • • • • • • • • • • • • • • • • •				
Only plates Will	in between JU LU	than 3 hours before 300 colonies counted	u, when		
of undilute	ed sample is pla	ted, colony density	may be 1	ess	
Only two signif	ficant figures r	ecorded and calculat	ed as	1000	
The first of the second s	lata aquat/ml				



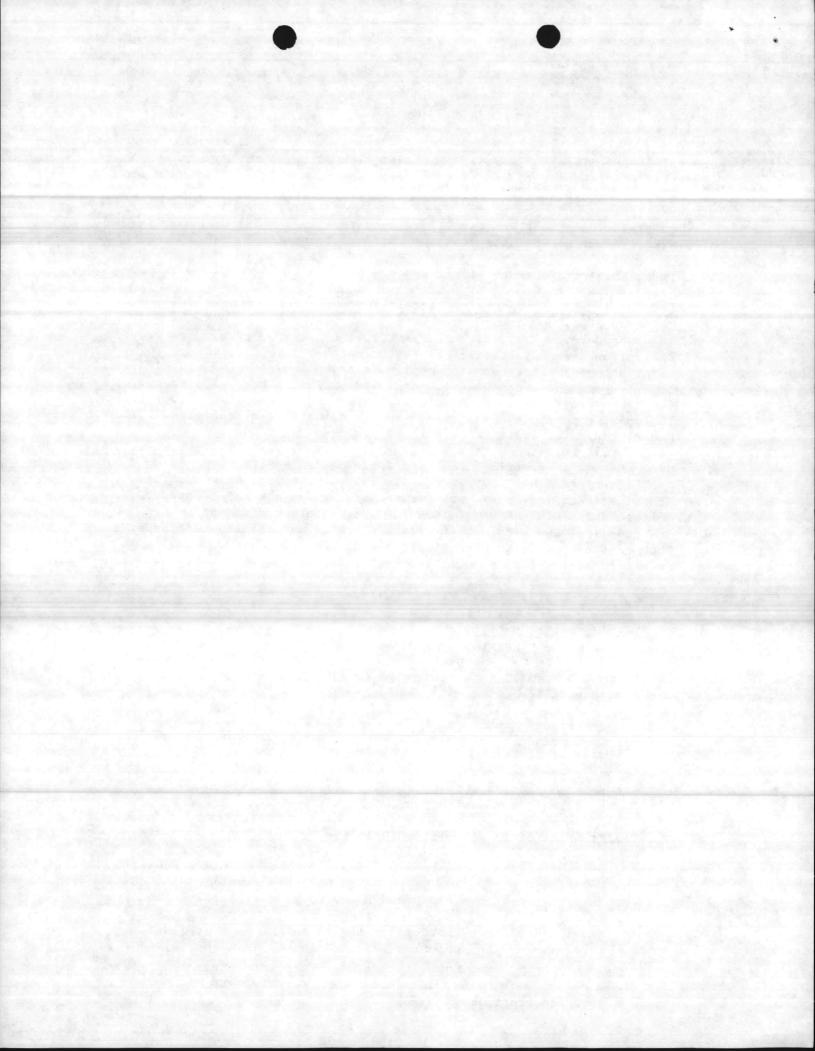
GENERAL LABORATORY PRACTICES (Continued)

10.	Levine's Eosin Methylené Blue Agar (EMB)						
	Manufacturer	Difco	Lot No.	701060	8/85		
	Medium composi Final pH 7.1 <u>+</u>	tion 37.5g per liter 0.2			<u>s</u> <u>s</u>		
11.	Sterility Test	Broth			<u>s</u>		
	Manufacturer	Difco (Tryptic Soy Broth)	Lot No	709765	8/87		



METHODOLOGY

M-Endo hr	gy specified in "Standard Methods" 14th edition, or EPA manual5 oth, M-Endo agar, or Les Endo agar used in a single
ste	p procedure
cat	urated absorbent had for 1.5 to 2 hours at 35 + 0.5 C; then on
M-F	ndo broth at Les Endo agar for 20 to 22 hours at 35 ± 0.5 CNA
H L	
	al Coliform Membrane Filter Procedure
Sam	ples containing excessive bacterial populations (greater than 200), confluency, or turbidity retested by the MPN
	procedure
Fil	tration assembly sterile at start of each series
Abs	orbent pads saturated with medium, excess discarded; or 4.0 ml of agar medium can be used per culture dish instead of a padS
C	of agar medium can be used per culture dish instead of a planting.
Tes	it sample portions measured and not less than 100 ml analyzedS anel rinsed at least twice with 20- to 30-ml portions of
Fur	sterile buffered waterS
MF	removed with sterile forceps, grasping outside effective
	filtering area
MF	rolled onto medium pad or agar so air bubbles are not trapped5
As	start and finish MF control test (rinse water, medium and
	supplies) run with each filtration series and results recorded S
Whe	en controls indicate contamination occurred, all data on S
	affected samples rejected and resampling requested
· a.	Incubation of Membrane Filter Cultures
	Total incubation time 22 to 24 hours at 35 \pm 0.5 C
	Incubated in tight-fitting culture dishes or loose-fitting
	dishes incubated in high relative humidity chambersS
b.	Membrane Filter Colony Counting
	Samples repeated when coliforms are "TNTC" or colony growth
	is confluent, possibly obscuring coliform development
	and/or detectionS
	Total coliform count calculated in density per 100 m1S
	Low power magnification device with fluorescent light
	positioned for maximum sheen visibilityS
c.	Verification of Total Coliform Colonies
	All typical coliform (sheen) colonies or at least five randomly
	selected sheen colonies from each positive sample verified
	in lauryl tryptose broth and BGLB
	Counts adjusted based on verificationS
	All atypical coliform (borderline sheen) colonies or at least
	randomly-selected colonies verified in LTB and BGLBS
	Counts adjusted based on verificationS
	Sheen colonies in mixed confluent growth reported and
	verified (optional)



METHODOLOGY (Continued)

d. MF Field Equipment

	Model		
Only standard laboratory MF	procedures adapted to field		

2. Total Coliform Most Probable Number Procedure

a. Presumptive Test

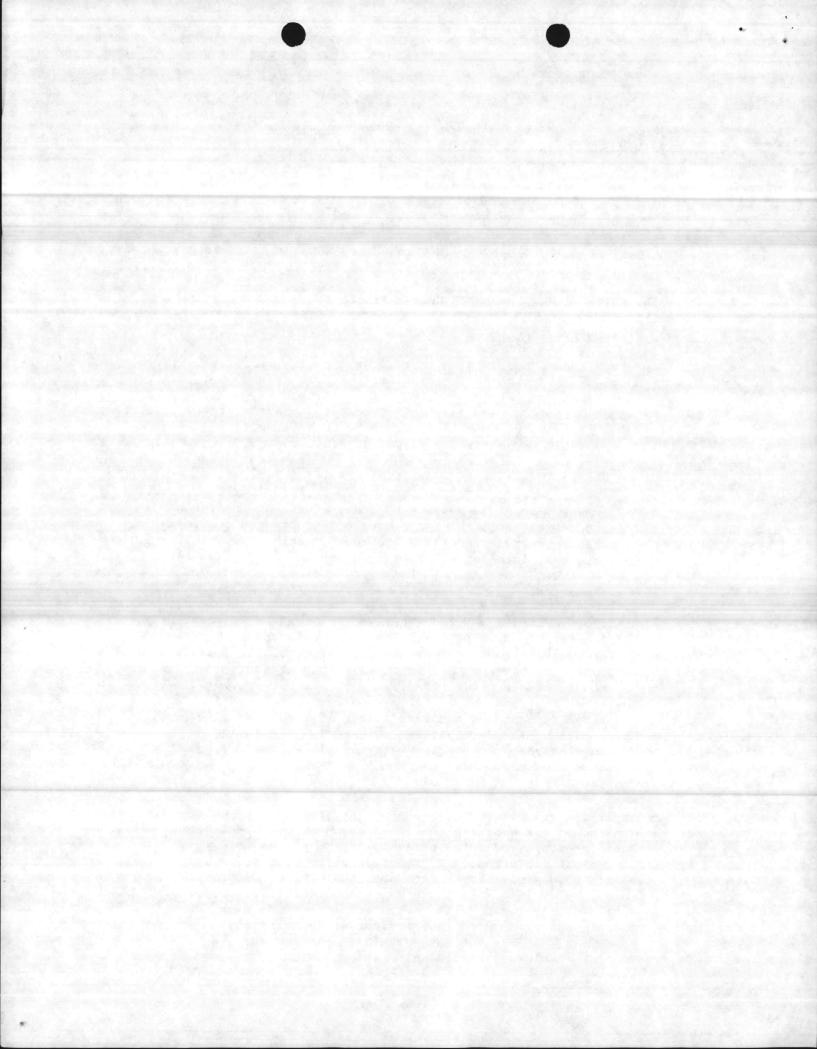
Five standard portions, either 10 or 100 mlS
Sample shaken vigorously immediately before testS
Tubes incubated at 35° + 0.5°C for 24 + 2 hoursS
Examined for gas (any gas bubble indicates positive test)S
Tubes that are gas-positive within 24 hours submitted
promptly to confirm test S
Negative tubes returned to incubator and examined for gas within 48 + 3 hours; positives submitted to confirm
test <u>S</u>
Public water supply samples with heavy growth and no gas production confirmed for presence of supressed coliforms S
Adjusted count reported based upon confirmationS Adequate test labeling and tube dilution coding (optional)

.b. Confirmed Test

Presumptive positive tube gently shaken or mixed by rotating S	,
One loopful or one dip of applicator transferred from presumptive tube to BGLBS	
Incubated at $35^{\circ} \pm 0.5^{\circ}$ C : checked within 24 hours ± 2 hours for gas productionS	
Positive confirmed tube results recorded; negative tubes reincubated and read within 48 + 3 hoursS	5
Confirmation procedure carried out every $\overline{3}$ months on one sample from each problem water supply	5

c. Completed Test

Applied to 10 percent of all positive samples each quarterS
Applied to all positive confirmed tubes in each test completed S
Positive confirmed tubes streaked on EMB plates for
colony isolationS
Plates adequately streaked to obtain discrete colonies
Incubated at $35^{\circ} \pm 0.5^{\circ}$ C for 24 ± 2 hoursS
Typical nucleated colonies, with or without sheen on EMB
plates selected for completed test identificationS
If typical colonies absent, atypical colonies selected
for completed test identificationS



METHODOLOGY (Continued)

c. Completed Test (Continued)

If no colonies or only colorless colonies appear, confirmed	
test for that particular tube considered negative	S
An isolated typical colony or two atypical colonies	
transferred to lauryl tryptose broth	S
Incubated at 35° + 0.5°C; checked for gas within 48 + 3 hours	
Cultures producing gas in lauryl tryptose broth within	
48 + 3 hours are considered coliforms	S

3. Analytical Quality Control

а.	A recor	d of analytical	quality control	tests available	
	for	review			S

Duplicate analyses

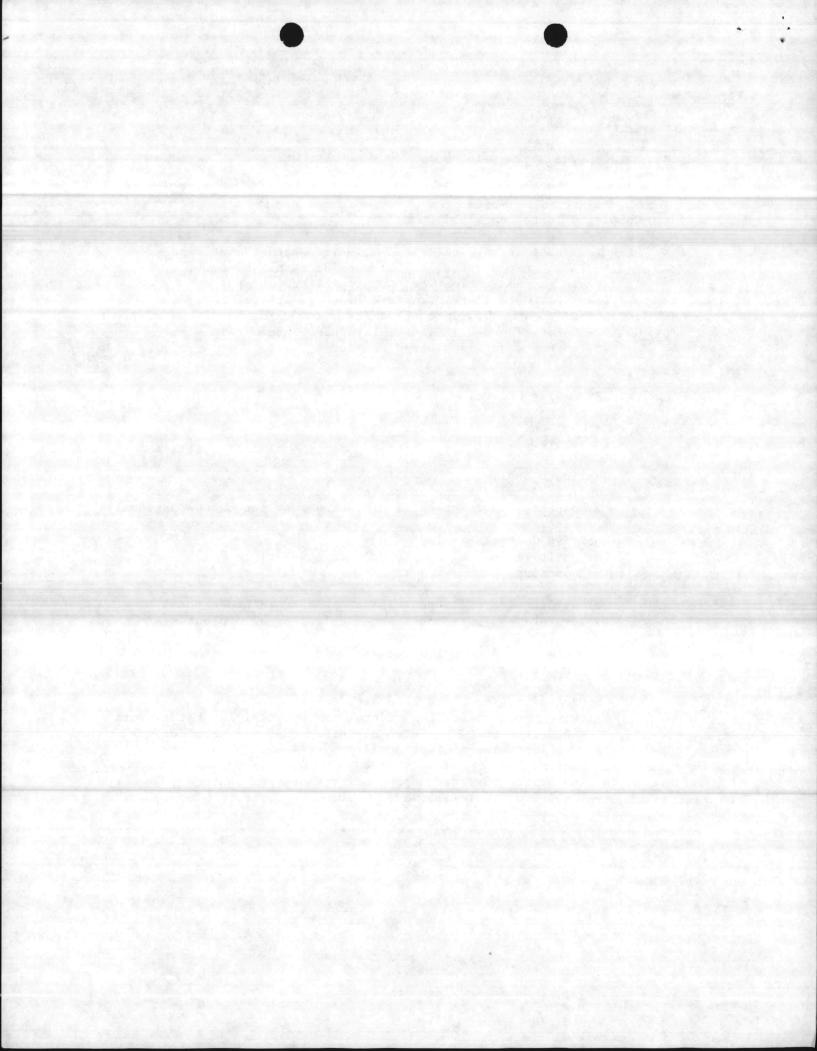
Duplicate analyses run on positive polluted samples not to exceed 10 percent but a minimum of one per month (optional)

Positive Control Samples

One positive control sample (polluted water) run each month (optional)

Colony Counting (If more than one Analyst in Laboratory) Two or more analysts count sheen colonies; all colonies are verified analysts' counts compared to verified counts; procedure is carried out at least once per month. (optional)

Check Analyses by State Laboratories A minimum of samples proportional to the local laboratory work load processed by State Laboratory (see criteria for recommendations) (optional)



SAMPLE COLLECTION, HANDLING, AND PRESERVATION

Representative samples of potable water distribution system	S
Minimal sampling frequency as specified in the National Interim	
Primary Drinking Water Regulations	S
Sample collector trained and approved as required by State regulatory	. 8
authority or its delegated representative	S

1. Sample Bottles

Sodium thiosulfate, (10 mg per 100 ml of sample) added to	
sample bottles before sterilization	S
Ample air space remains after sample collected to allow for	
adequate mixing	S

2. Sampling

Sample collected after maintaining a steady flow for 2 to 3 min
to clear service lineS
Tap free of aerator, strainer, hose attachment, water
purification, or other devicesS
Samples refrigerated when possible during transit and storage
periods in the laboratory (optional)

3. Sample Identification

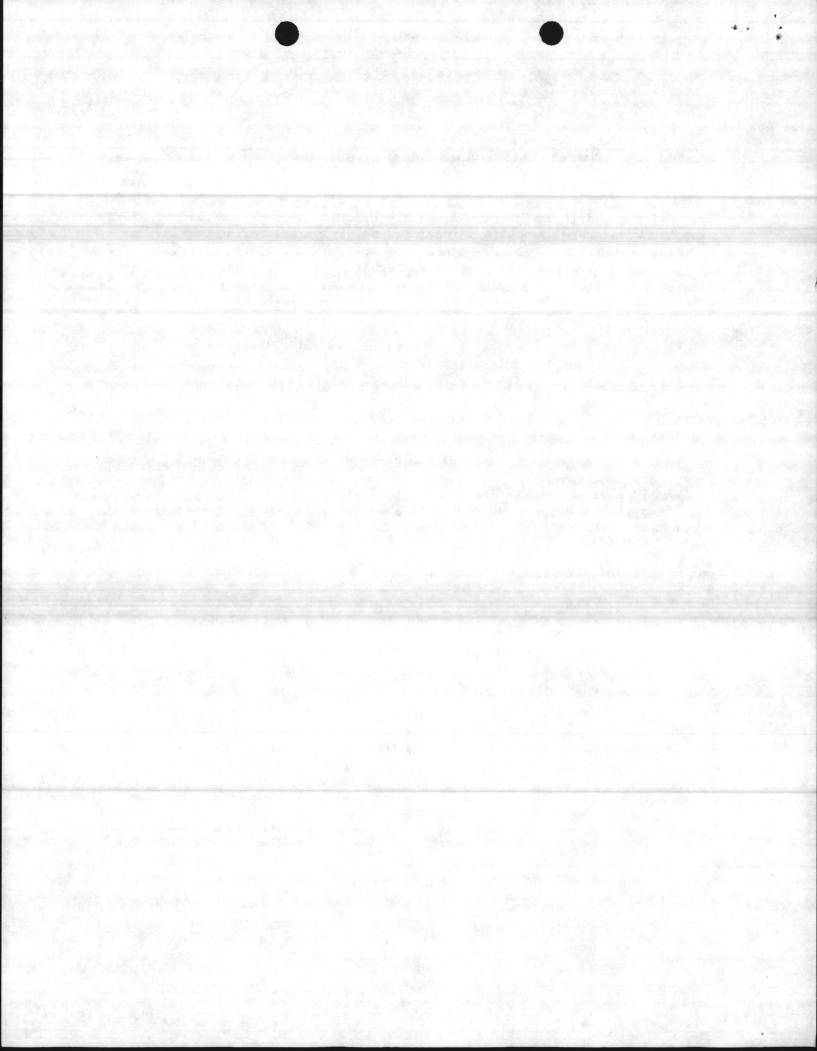
Sample identified immediately after collectionS
Identification includes, water source, location, water supply
identification number, time and date of collection, and
collector's name; insufficiently identified samples
discardedS
Chlorine residual where applicableS

4. Sample Transit Time

Transit time for potable water samples sent by mail or
commercial transportation, not in excess of 30 hours
No sample processed after 48-hour transit/storageS
Samples delivered to laboratory by collectors examined the
day of collectionS
Data marked as questionable on samples analyzed after 30 hours

5. Sample Receipt in Laboratory

Sample logged in when received in laboratory, including date and
time of arrival and analysisS
Chain-of-custody procedures required by State regulations followed S



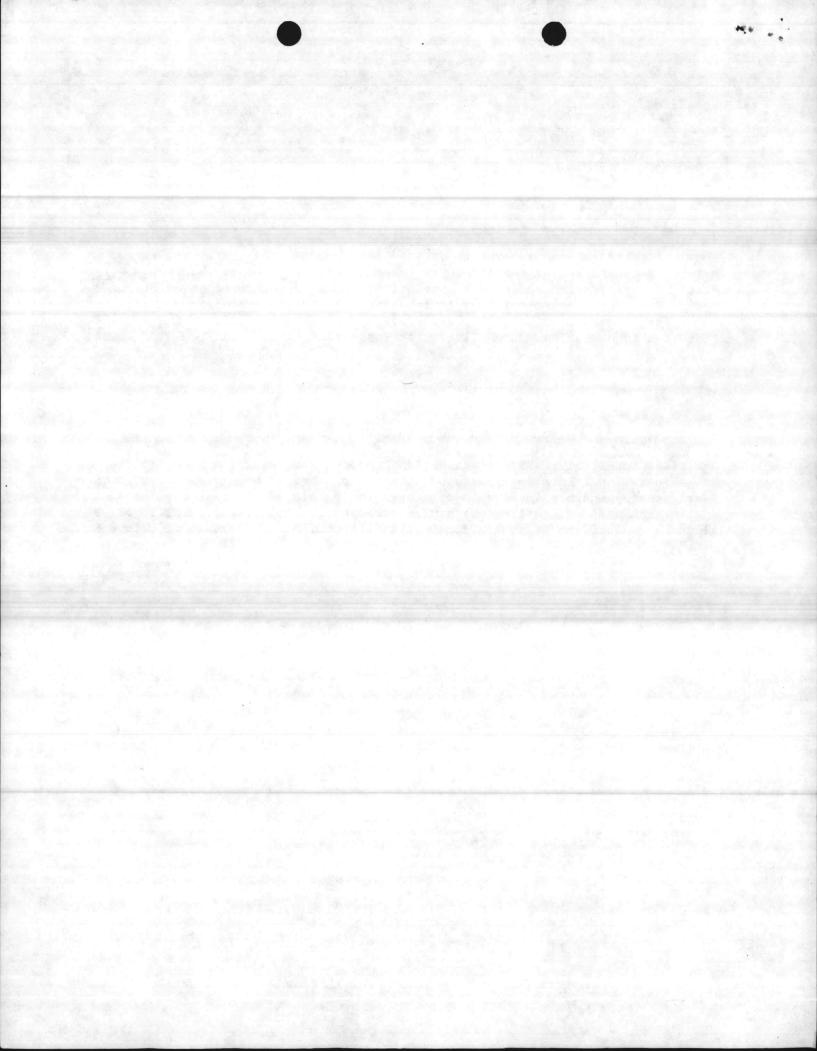
DATA REPORTING

Sample information and laboratory data fully recorded......S Direct MF counts and/or confirmed MPN results reported promptly.....S After MF verification and/or MPN completion, adjusted counts reported.....S One copy of report form retained in laboratory or by State program for 3 years.....S

Test results assembled and available for inspection (optional)

ACTION RESPONSE TO LABORATORY RESULTS

Unsatisfactory test results given action response and resampled as defined	
in National Interim Primary Drinking Water Regulations	5
State and responsible local authority notified within 48 hours after	
check samples confirm coliform occurrence	5
All data reported to State and local authorities within 40 days S	3



Ronald H. Levine, M.D., M.P.H. STATE HEALTH DIRECTOR

DIVISION OF HEALTH SERVICES STATE LABORATORY OF PUBLIC HEALTH 306 N. Wilmington St. P.O. Box 28047 Raleigh, N.C. 27611-8047

March 20, 1984

Commanding General Marine Corps Base Camp LeJeune, NC 28542

Attn: Facilities - NREAD Elizabeth Betz

Dear Ms. Betz:

It has been several months since I visited your laboratory to conduct a certification evaluation. We have not yet received a letter advising us of steps being taken to correct the deviations.

I hope this is just an oversight and that you are not encountering problems. Please let me hear from you soon so that we can send your certificate.

Thank you very much.

Sincerely,

E. D. Beesley Laboratory Certification Evaluator

GOVERNOR

EDB/hw

Ronald H. Levine, M.D., M.P.H. STATE HEALTH DIRECTOR

Paleta 1. C. C. Maria P.

Ronald H. Levine, M.D., M.P.H. STATE HEALTH DIRECTOR

DIVISION OF HEALTH SERVICES STATE LABORATORY OF PUBLIC HEALTH 306 N. Wilmington St. P.O. Box 28047 Raleigh, N.C. 27611-8047

September 26, 1983

Commanding General Marine Corps Base Camp LeJeune, NC 28542

ATTN: Facilities - NREAD Elizabeth Betz

Dear Ms. Betz:

Enclosed is a copy of the narrative report based on the survey of your laboratory for certification to analyze drinking water samples for EPA compliance. Please send a letter explaining what has been done to correct each of the deviations. The list of corrections should be sent to the above address. This laboratory is accredited for 60 days pending receipt of the above mentioned letter.

Upon receipt of acceptable corrections, we will continue your interim certification to the date shown on your certificate.

If you have any questions regarding certification or if we can be of assistance in any other way, please contact us.

Sincerely,

E.D. Beesley

E. D. Beesley Hur-Laboratory Certification Evaluator

EDB/sy

Enclosure

the state of the second se E.D. Seesley

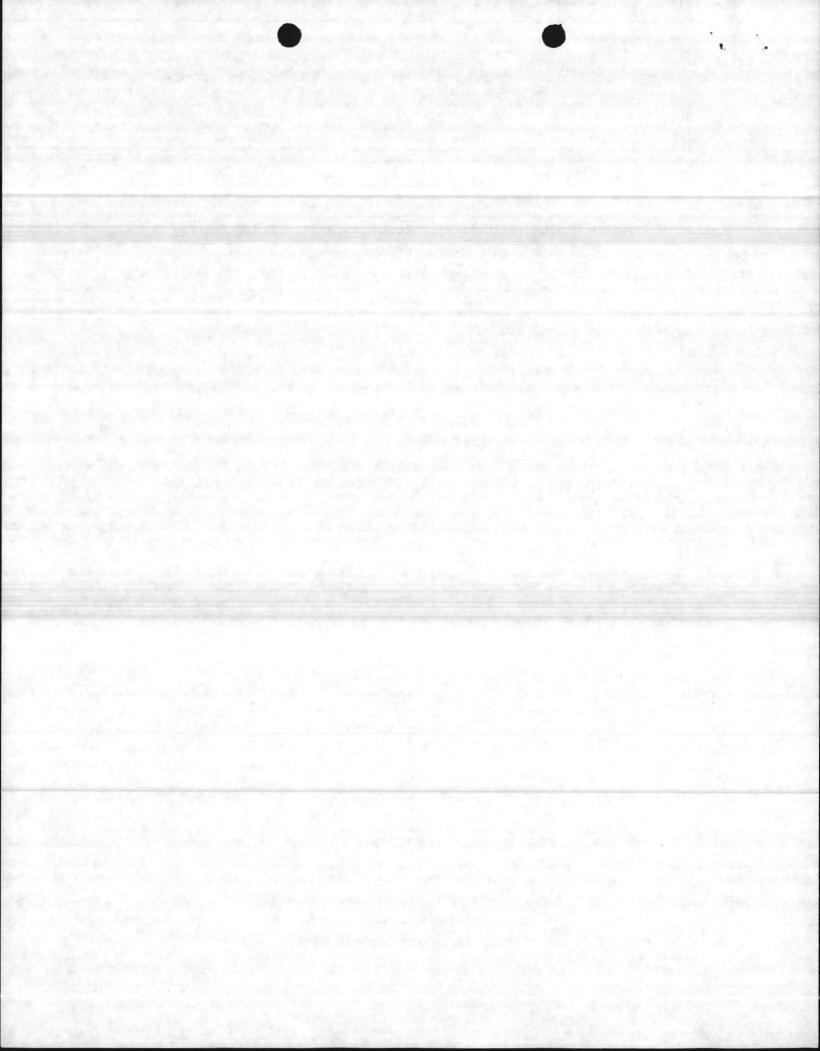
REPORT OF AN ON-SITE EVALUATION QUALITY CONTROL LABORATORY BACTERIOLOGY LABORATORY ENVIRONMENTAL BRANCH, NATURAL RESOURCES AND ENVIRONMENTAL AFFAIRS DIVISION BASE MAINTENANCE DIVISION, MCB CAMP LEJEUNE CAMP LEJEUNE, NORTH CAROLINA

SEPTEMBER 8, 1983

BY:

E. D. BEESLEY LABORATORY CERTIFICATION EVALUATOR ENVIRONMENTAL SCIENCES BRANCH

LABORATORY SECTION NORTH CAROLINA DIVISION OF HEALTH SERVICES NORTH WILMINGTON STREET RALEIGH, NORTH CAROLINA 27611



QUALITY CONTROL LABORATORY CAMP LEJEUNE, N. C. SEPTEMBER 8, 1983

I. INTRODUCTION

The equipment and procedures employed in the bacteriological analyses of water by this laboratory conformed with the provisions of the North Carolina Safe Drinking Water Regulations, except for the items indicated.

II. DEVIATIONS AND RECOMMENDATIONS

- A. Methodology
 - 1. Analytical Quality Control

In order to maintain certification for MPN a known positive sample must be tested and carried through to completion at least once per quarter (a good practice is to use the same sample for both the MPN and MF quality control.)

Results must be recorded in the quality control log.

III. LIST OF PERSONNEL

NAME

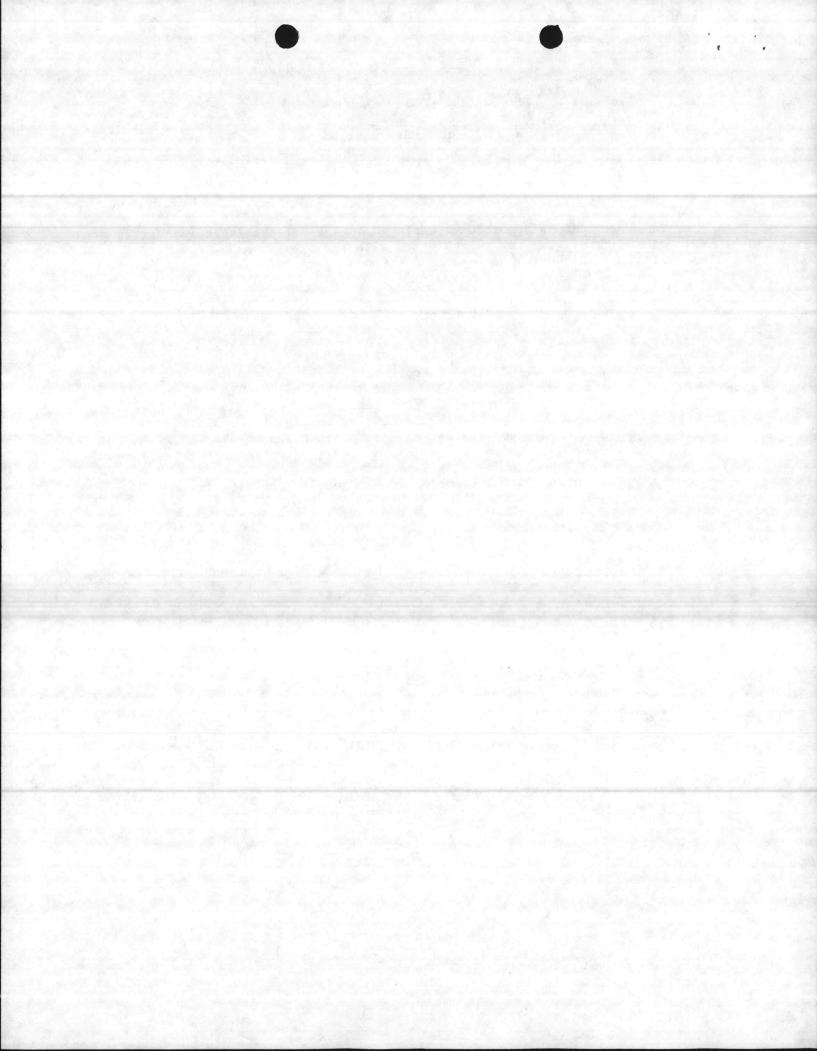
POSITION

TEST NORMALLY PERFORMED

Elizabeth A. Betz	Supervisor	MF & MPN
Hoy Burns	Technician/Analyst	MF & MPN
Bob Lachapelle	Technician/Analyst	MF
Gaines Honeycutt	Technician/Analyst	MF
Gerald Monahan	Technician/Analyst	MF
	rechnician/Analyst	MF

IV. CONCLUSION

The procedures and equipment in use at the time of this survey were in general compliance with the provisions of the North Carolina Drinking Water Regulations (10NCAC 9D .0301 - .0330). This laboratory is accredited for 60 days pending correction of the deviations and receipt of a letter by the evaluation officer detailing the corrections made. Upon receipt of such a letter, full accreditation will be given.



STATE LABORATORY OF PUBLIC HEALTH

DIVISION OF HEALTH SERVICES

NORTH CAROLINA DEPARTMENT OF HUMAN RESOURCES

P. O. BOX 28047, 306 NORTH WILMINGTON STREET,

RALEIGH, NORTH CAROLINA 27611

FORMS FOR ON-SITE EVALUATION OF LABORATORIES INVOLVED IN ANALYSIS OF PUBLIC WATER SUPPLIES

MICROBIOLOGY

LAB I.D. NUMBER: 37807

LABORATORY: <u>Camp</u> STREET:	LeJeune Quality Natural Resourc Commanding Gen Marine Corps B	es and Env	ironmental Affair	enmental Branch, s Division ilities - NREAD
CITY:	Camp LeJeune	STATE :	North Carolina	28542
TELEPHONE NUMBER:	919-451-5977			
SURVEY BY:	E. D. Beesley			
AFFILIATION:	North Carolina	a Division	of Health Service	es

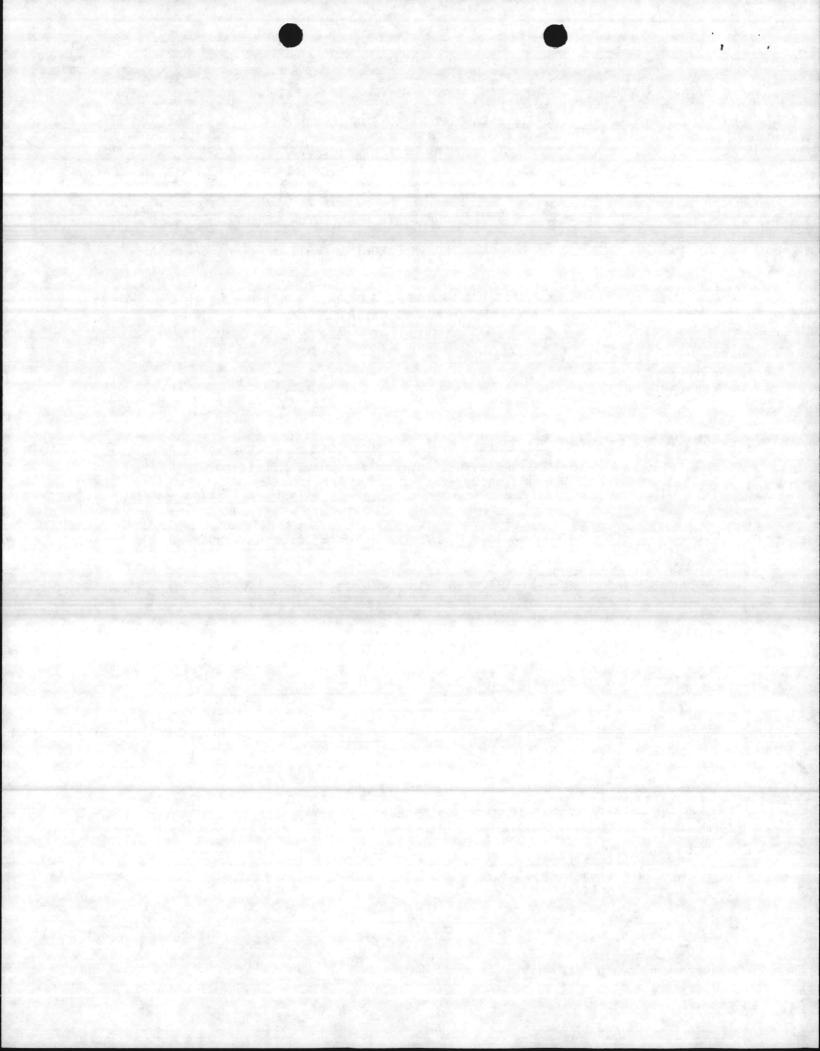
Codes for Marking On-Site Evaluation Forms:

September 8, 1983

S - Satisfactory X - Unsatisfactory NA - Not Applicable

DATE:

DHS FORM 2907 Revised (11/82) Laboratory



PERSONNEL

	HS	BA/BS	MA/MS	PH.D	METHOD(S)	(YEARS/AREA)
		100000				(ILARO/AREA
abeth A. Betz*	x	BS Chem.			MF-MPN	4 yrs.
Burns Lachapelle es Honeycutt Ld Monahan	X X X X	lyear AAS**	*** **	ence	MF–MPN MF MF MF	7 yrs. 2 yrs. 2 yrs. 1 yr.
	Surns Lachapelle 25 Honeycutt	Burns X Lachapelle X es Honeycutt X	Chem. Chem. Surns X lyear Lachapelle X lyear es Honeycutt X AAS**	Chem. Chem. Surns X lyear** Lachapelle X lyear*** es Honeycutt X AAS****	Chem. Chem. Surns X lyear** Lachapelle X lyear*** AAS****	Chem. Burns X lyear** MF-MPN Lachapelle X lyear*** MF es Honeycutt X AAS**** MF

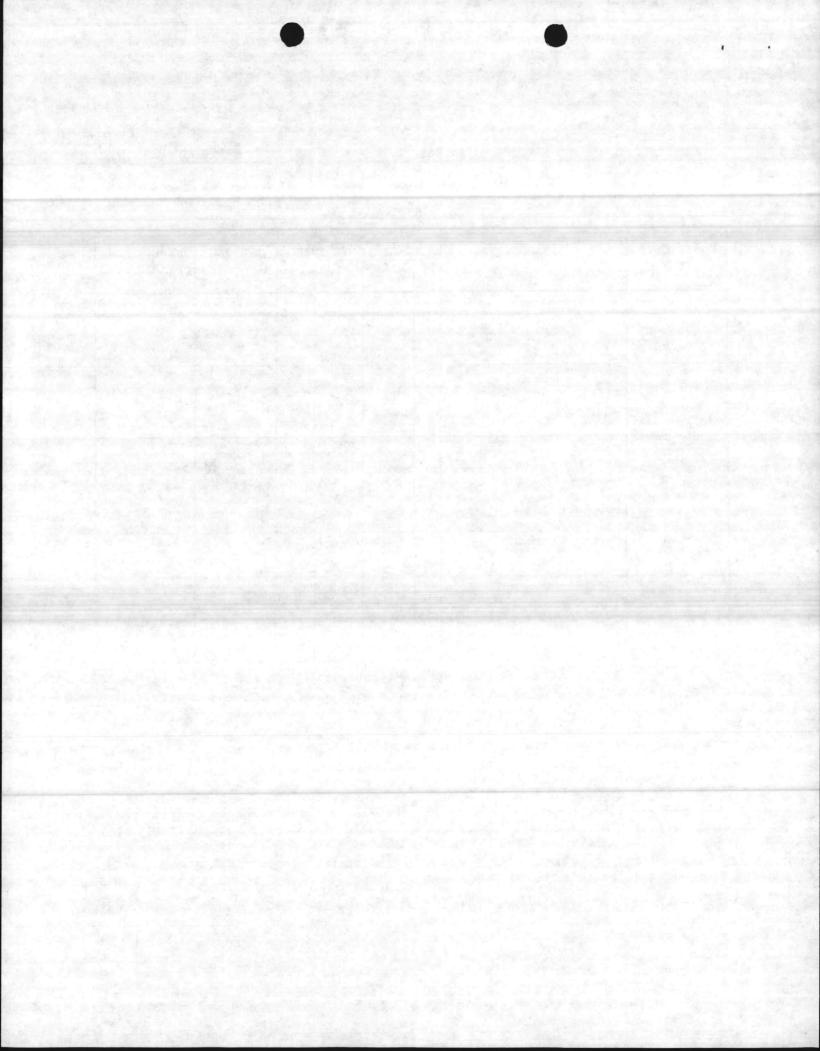
* Attended NCDHS Water Microbiology Workshop
** 14 Mo. Navy Clinical Lab School, Reg. Med. Tech.

*** 14 Mo. " " " ", Lab Supervisor 4 years **** 6 Yrs. N. C. Dept. of Natural Resources LABORATORY FACILITIES

Space in laboratory and preparation room is adequate for needs during peak work periods (200 ft and 6 linear ft. of usable bench space per analyst).

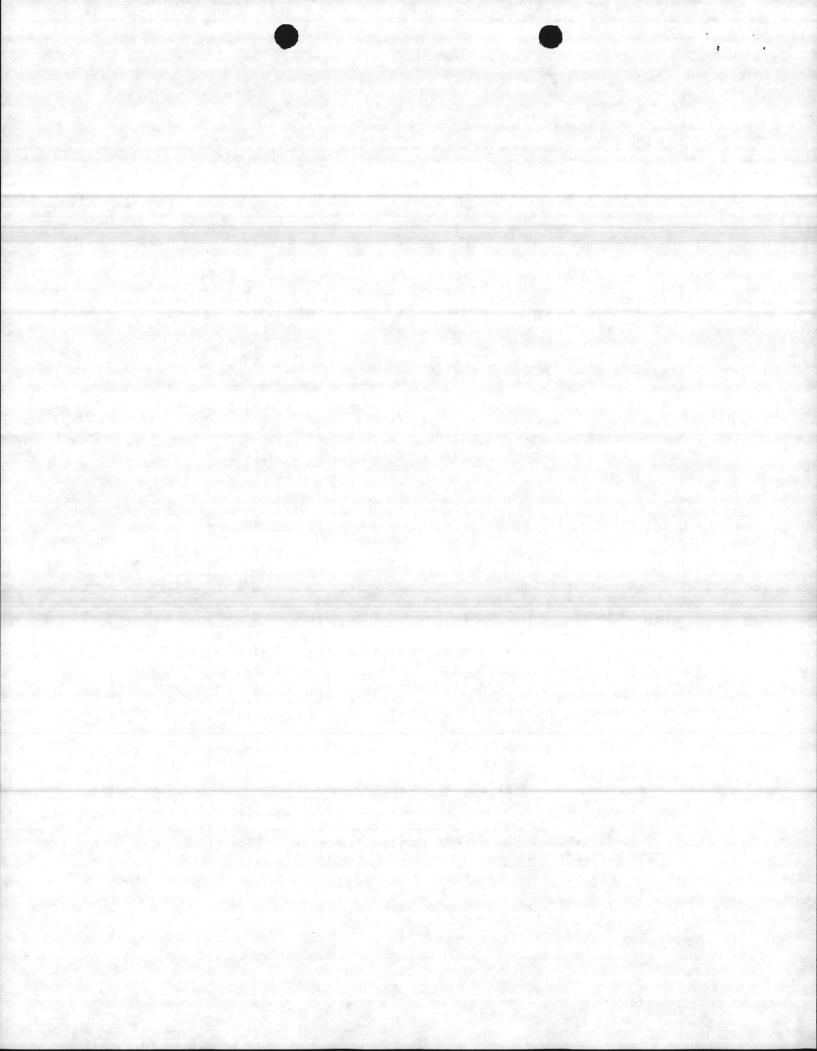
Facilities are clean, with adequate lighting (100 ft-candles) and air-conditioning.

Satisfactory



LABORATORY EQUIPMENT, SUPPLIES, AND MATERIALS

		Fisher	750
Ma	anufacturer	Orion	Model 701
A	liquot of star	dard pH 7.0 buffer used	use period; record maintained <u>s</u> only once <u>s</u> n opened <u>s</u>
Ba	alance-Top Loa	der or Pan	
. Ma	anufacturer	Ohaus	Model <u>Harvard Trip</u>
Ca	preparation alibrated annu	of 2-g quantities)	ely (for a general media
Th	nermometers		
Ce	ertified Therm	ometer:	
Ma	nufacturer	Fisher (Kessler)	Certificate # 836-212
Ma	ximum Registe	ring:	
Ma	nufacturer	Brooklyn	
Le	thermometer checked qua gible graduat	rterly	against a certified ccuracy; metal thermometers
In	cubator or In	cubator Room	
Ma	nufacturer	Precision	ModelMZ
Su Th	ermometer grad liquid and	duated in 0.5°C increment located on top and botto	ts with bulb immersed in m shelvess yes in all areas used
	ifamm hammen		



LABORATORY EQUIPMENT, SUPPLIES, AND MATERIALS (Continued)

5. Autoclave

6.

7.

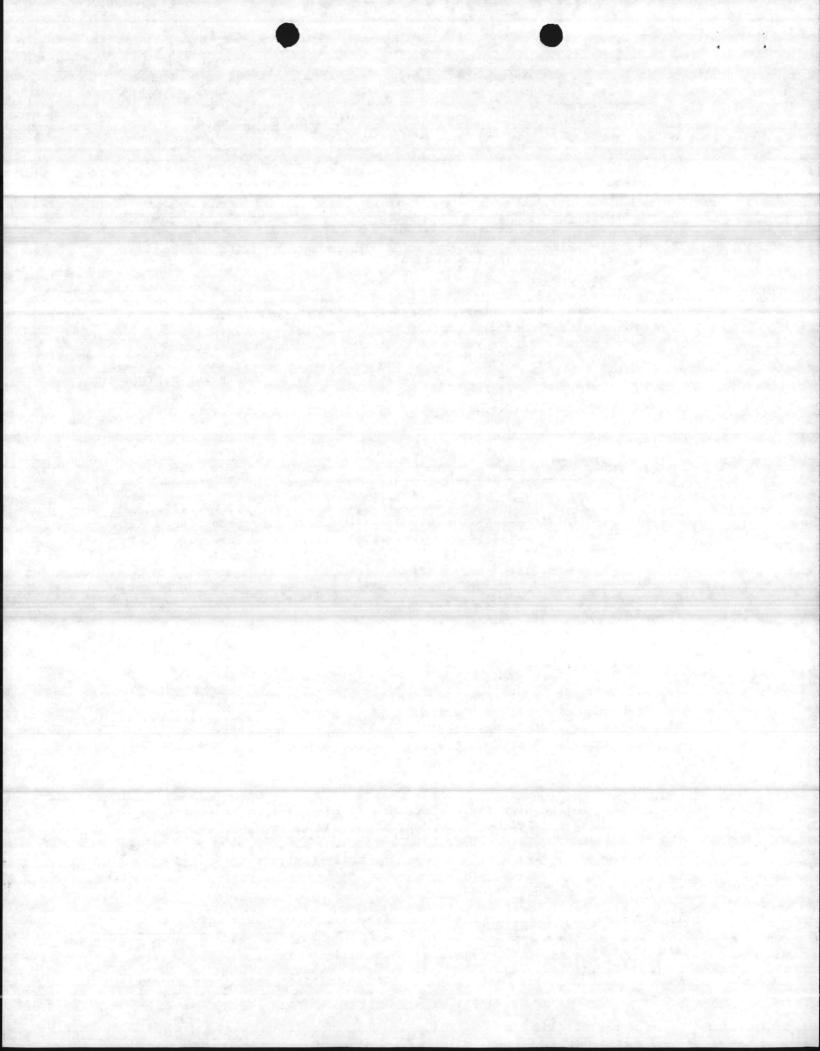
8.

9.

10.

Manufacturer <u>Market Forge</u>	Model Sterilmatic
Reaches sterilization temperature (121°C), sterilization cycle, and requires no mo	re than 45 min. for a
complete cycle Temperature checked at least weekly with a registering thermometer calibrated in 19	maximum
and results recorded Pressure and temperature gauges on exhaust a	····· S
safety valve	
No air bubbles produced in fermentation via Record maintained on time and temperature for	or each sterilization
cycle	•••••••• <u> </u>
Hot-Air Oven	
Manufacturer NA	Model
Operates at a minimum of 170°C Thermometer inserted or oven equipped with a thermometer device Time and temperature record maintained for e	temperature-recording
Thermometer bulb in sand (optional)	each sterilization cycle
Refrigerator	
Temperature maintained at 1° to 5.0°C	<u>s</u>
Inoculation Equipment	
Sterilized loops of at least 3-mm, diameter,	, 22 to 24 gauge
Nichrome, Chromel, or platinum-iridium w Disposable, dry heat-sterilized, hardwood ap presterilized loops	oplicator sticks or
Optical Equipment	<u>NA</u>
Low power magnification device (preferably b with 10 to 15X) with fluorescent light s	source for counting
MF colonies Colonies counted with a mechanical hand tall	y (optional)
Membrane Filtration Equipment	
Manufacturer Millipore	Model _
Made of stainless steel, glass, or autoclava	ble plastic
Nonleaking and uncorroded	prostic

4



LABORATORY EQUIPMENT, SUPPLIES, AND MATERIALS (Continued)

11. Membrane Filters and Pads

Manufacturer	Millipore	Type HAWG
이 같은 것 같은		

Filters recommended by manufacturer for water analyses..... Filters and pads presterilized or autoclavable..... Lot numbers and dates of receipt of membrane filters recorded (optional)

12. Glass, Plastic, and Metal Utensils for Media Preparation

Washing process provides glassware free of toxic residue as demonstrated by the inhibitory residue test and results recorded......S

Detergent: DISPERSE (American Scientific)

Glass items of borosilicate, free of chips and cracks......<u>S</u> Utensils clean and free from foreign residues or dried medium.....<u>S</u> Plastic items clear with visible graduations......S

13. Sample Bottles

Wide-mouth hard glass bottles; stoppered or <u>plastic screw-capped</u> ;
capacity at least 120 mlS
Glass-stoppered bottles with tops covered with aluminum foil or kraft paperÑA
Whirl-Pak Bags
Screw-caps have leakproof nontoxic liners that can withstand repeated sterilization (30 min at 121°C)S
Sterility of each batch of sample containers checked using non- selective broth and results recordedS

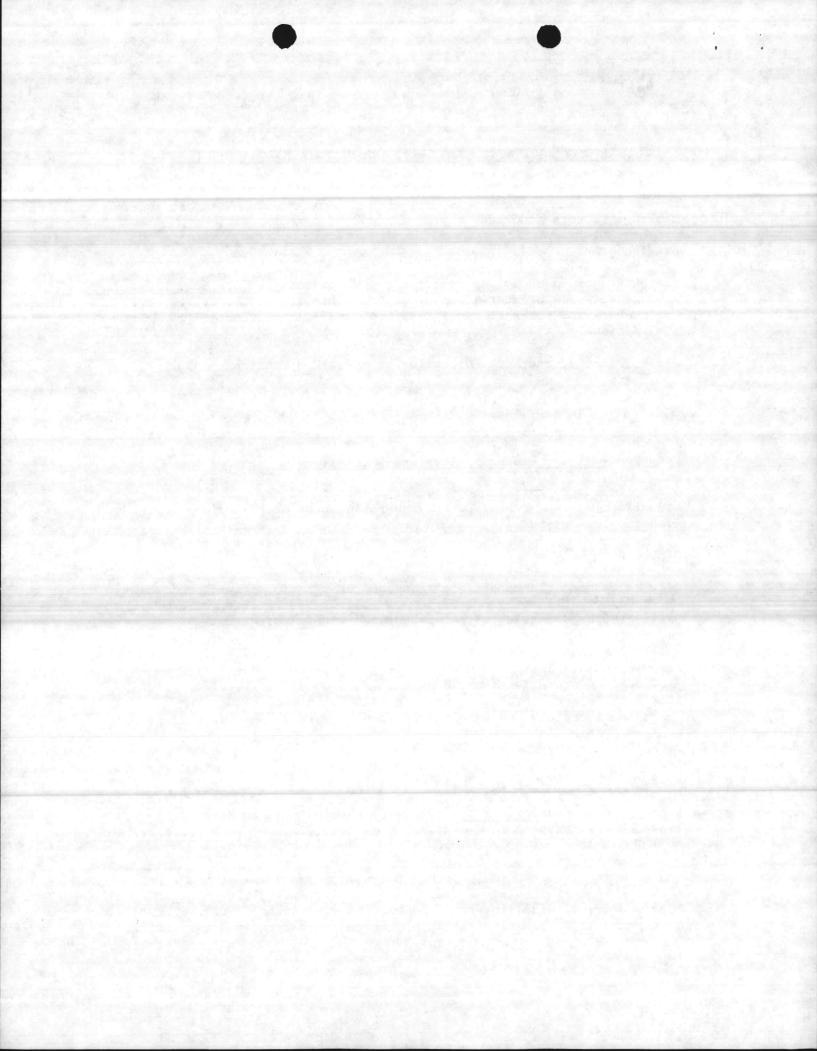
14. Pipets

Brand	Falcon	Туре
·		

Sterile;	glass o	or plastic;	with a	2.5	percent	tolerand	 ·····S
Tips unbr	oken; gi	raduations	distinctl	y ma	arked		 S

15. Pipet Containers

Aluminum or stainless steel.....NA Pipets wrapped in quality kraft paper (char-resistant)......NA Open packs of disposable sterile pipets resealed between uses......S



LABORATORY EQUIPMENT, SUPPLIES, AND MATERIALS (Continued)

16. Culture Dishes

Brand	Millipore Pvrex	49 X 9 Type 100 X 15	
Drund	Pyrex	Type _100) X 15

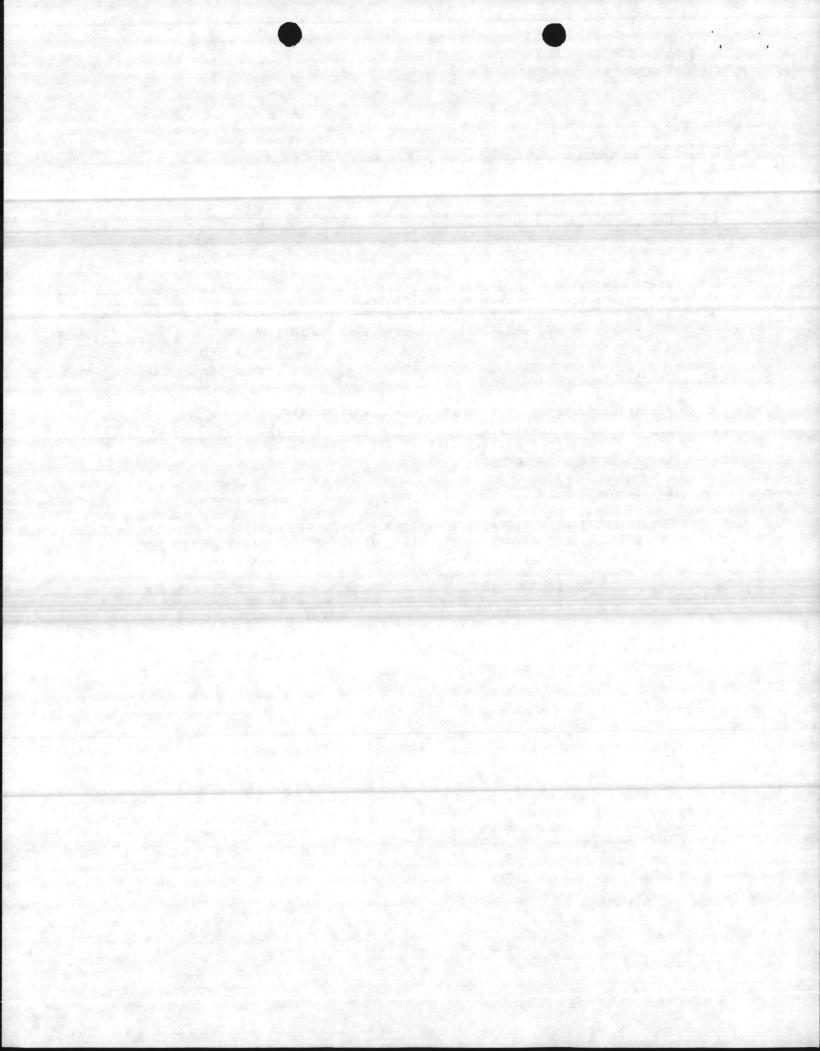
Sterile <u>plastic</u> or <u>glass</u>.....<u>S</u> Open packs of disposable sterile plastic dishes resealed between uses.<u>S</u> Dishes are in containers of aluminum or stainless-steel with covers or are wrapped with heavy aluminum foil or char-resistant paper.....S

17. Culture Tubes and Closures

Sufficient size to contain medium and sample without danger of
spillageS
Metal or plastic capsS
Borosilicate glass or other corrosion-resistant glassS

18. Maintenance

Ser	vice cont	racts or ap	pproved	interna	al prot	ocol main	ntained (on	
	balance,	autoclave,	, water	still,	etc.;	service	records	entered	
	in a log	book							. S



GENERAL LABORATORY PRACTICES

1. Sterilization Procedures

Timing for sterilization begins when autoclave reaches 121°C.....<u>S</u> Tubed broth media and reagents sterilized at 121°C 12 to 15 min.....<u>S</u> Tubes and flasks packed loosely in baskets or racks for uniform

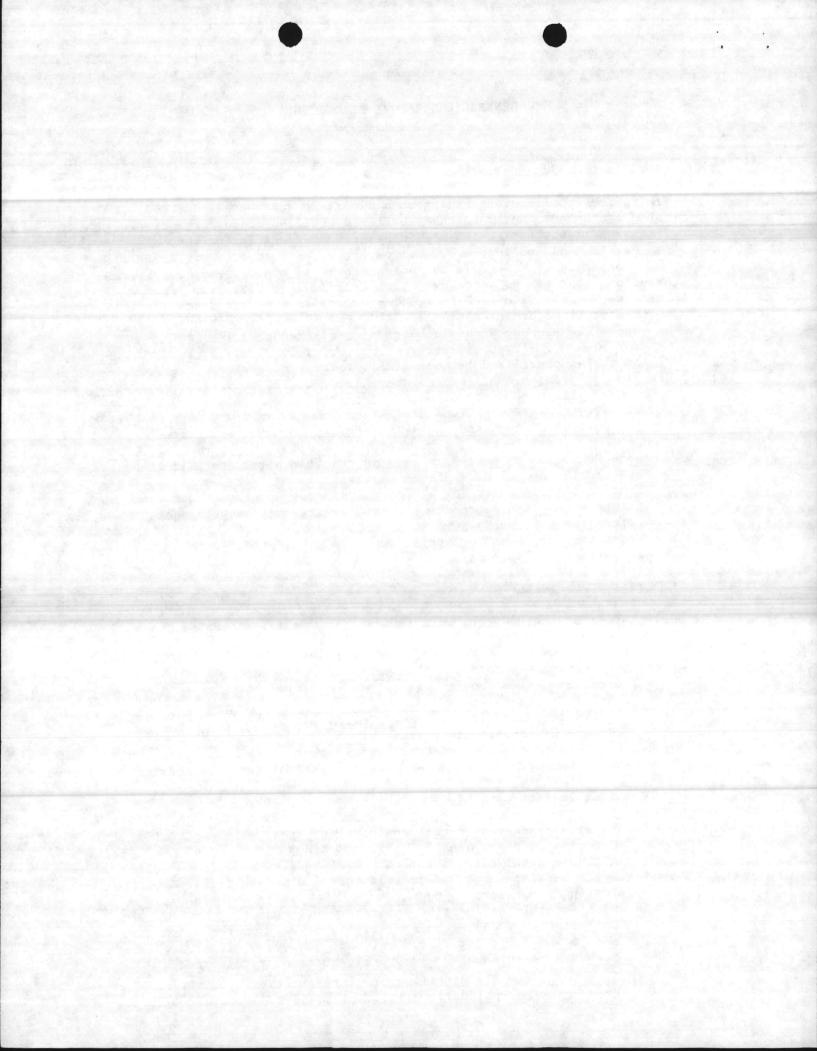
at 170°C for 2 hours......S MPN media removed and cooled as soon as possible after sterilization and stored in cool dark place (optional)

UV light or boiling water for at least 2 min may be used on membrane filter assemblies to reduce bacterial carry-over between each filtration (optional)

Heat-sensitive tapes and/or strips/ampoules used during sterilization (optional)

2. Laboratory Pure Water

Only laboratory pure water, used in preparing media, reagents, rinse
water, and difution water.
Laboratory pure water not in contact with heavy metals.
Source: Laboratory-prepared V Purchased
If Laboratory-prepared:
Still Manufacturer Consistenty
Still Manufacturer Corning Megapure 6L and 3L
Deionizer Manufacturer <u>Corning High Cap</u> Record of recharge frequency
internet of recharge frequency
Production rate and quality adequate for laboratory needsS
Inspected, repaired, cleaned by service contract or in-house service. S
a. Chemical Quality Control
Record of satisfactory annual analyses for trace metals (Cadmium,
Chromium, Copper, Nickel, Lead, and Zinc)
A single motal not areator the 0.05 (1
A single metal not greater than 0.05 mg/1
rotar metars. equal to or less than 1.0 mg/1
Testing laboratory NCDHS Date (or
Record of monthly analyses of laboratory pure water
Conductance: >0.5 megohm resistivity or <2.0 micrombos/cm
pH: 5.5 - 7.5
Standard plate count: <10,000/ml. Stored or deionized;
<1000 for 11 the state of action of delonized;
<1000, freshly distilled or ultra-pure



GENERAL LABORATORY PRACTICES (Continued)

b. Microbiological Quality Control

Test for bactericidal properties of	f distilled water
(0.8 - 3.0) performed at least	annually
Testing laboratory	Date Ratio

3. Rinse and Dilution Water

Stock buffer solution prepared according to "Standard Methods",
14th edition, or EPA Methods Manual S
Stock buffer solution adjusted to pH 7.2
Stock buffer autoclaved at 121°C, stored at 1° to 5.0°C
or filter sterilized <u>S</u>
Stock buffer labeled and datedS
Stock potassium phosphate buffer solution (1.25 ml) added per
liter distilled water for rinse and dilution water
Final pH 7.2 ± 0.1
MgSO ₄ MgCl ₂ 5 ml stock solution per liter

Media

Dehydrated media bottles kept tightly closed and protected from

dust and excessive humidity in storage areasS
Dehydrated media not used if discolored or cakedS
Laboratory pure water used in media preparationS
Media completely dissolved before dispensing to culture
tubes or bottles
MPN tube media with loose-fitting caps used in less than 1 week
Tube media in screw-capped tubes held no longer than 3 months
Media stored at low temperatures are incubated overnight prior
to use and tubes with air bubbles discardedS
Media protected from sunlight
ME modia stored in refrigereters last 11 11 1 11 1 1

MF media stored in refrigerator; broth media used within 96 hours, agar within two weeks if prepared in tight-fitting dishes.....S Ampouled media stored at 1° to 5.0°C and time limited to

manufacturer's expiration date.....

5. Quality Control of Media and Reagents

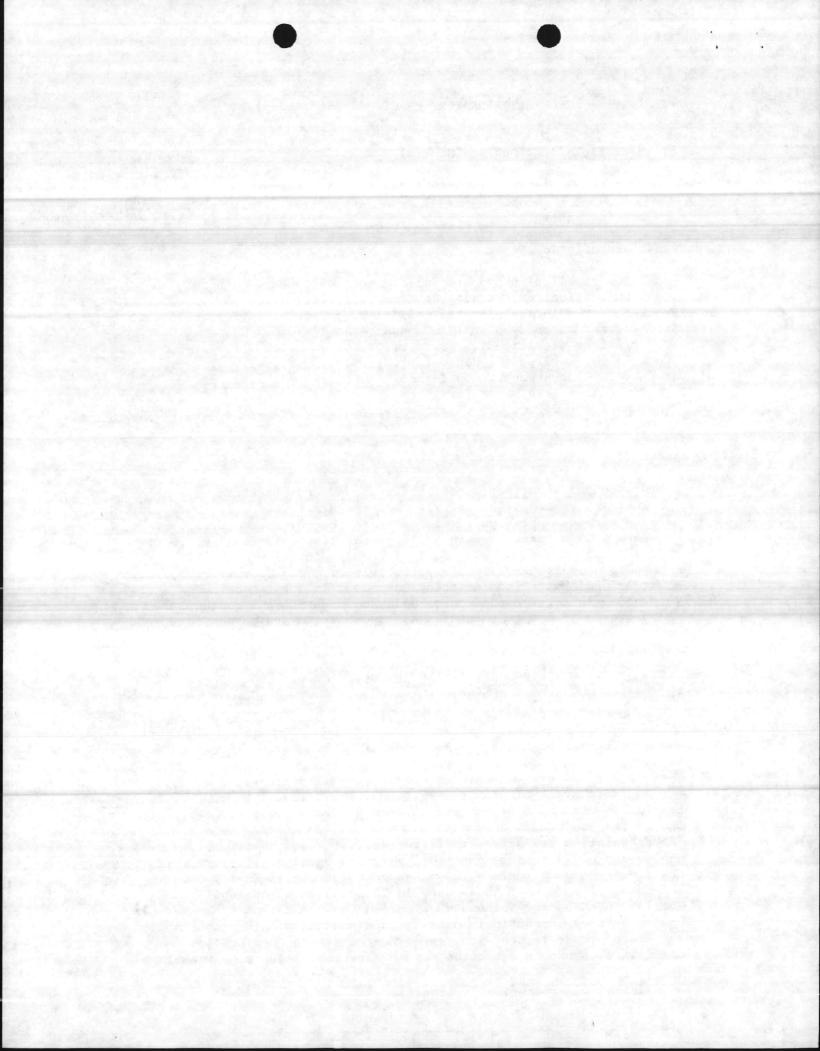
Satisfactory records containing complete quality control checks

on media ava	ilable for inspectionS
Laboratory chemi	cals of Analytical Reagent Grade
pH checked and r	ecorded on each batch of medium after
preparation	and after sterilization S
Causes for devia	tions beyond + 0.2 pH units specified
Media ordered on	a basis of 12-month need; purchases in 1/4 1b.
quantities,	except those used in large amounts (optional)

Bottles dated on receipt and when opened (optional) Opened bottles of routinely used media discarded within 6 months (if

stored in desiccator storage may be extended) (optional) Shelf life of unopened bottles not in excess of 2 years (optional)

New lots of media quality tested against satisfactory lot using natural water samples (optional)



GENERAL LABORATORY PRACTICES (Continued)

6. Lauryl Tryptose Broth

Manufacturer	Difco	

Lot No. 707667 8/87

Single Strength composition, 35.6g per liter pure water......S Single strength pH 6.8 ± 0.2; double strength pH 6.7 ± 0.2.....S Not less than 10 ml per tube......S Media made to result in single strength after addition of sample portions.....S

7. Brilliant Green Lactose Bile Broth

Manufacturer_	Difco		L	ot No.708037	5/87
Medium compos	ition 40g per	liter pure	water		<u>s</u>
Final ph 7.2	F U.Z				S

M-Endo Media

8.

9.

Manufacturer BBL

Lot No.15DMSX 9/86

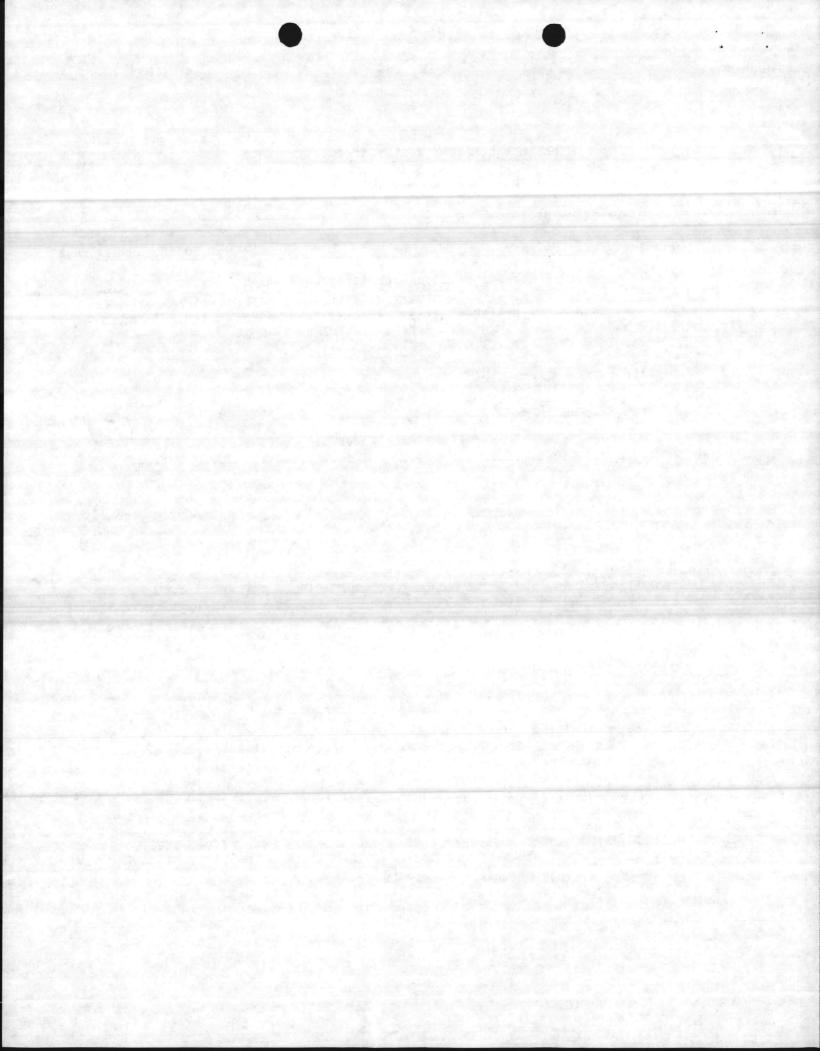
Medium composition 48.0g per liter pure water; optionally
15g agar added/1S
Reconstituted in laboratory pure water containing 2 percent ethanol (not denatured)
Final pH $7.2 + 0.2$
Medium held in boiling water bath until completely dissolved

Standard Plate Count Agar

Manufacturer Difco

Lot No.677117

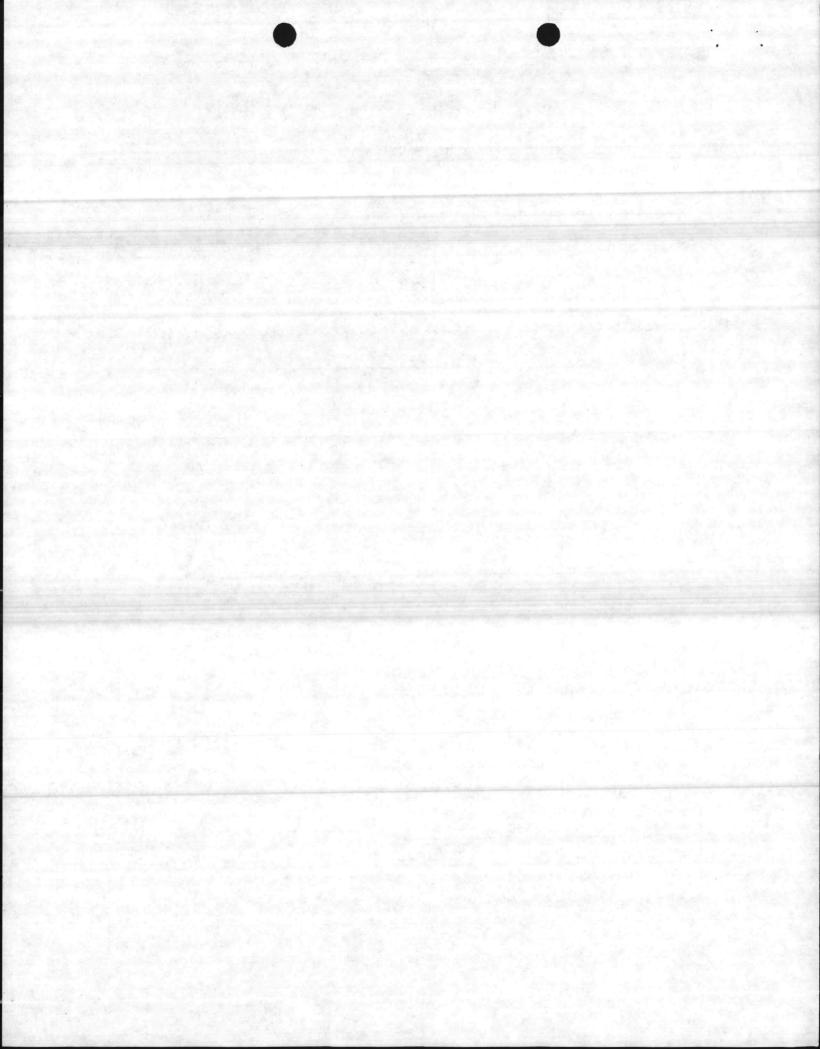
Correct composition, sterile and pH 7.0 + 0.2
Sterile medium not remeited a second time after sterilization
Culture dishes incubated 48 hours at $35^{\circ} \pm 0.5^{\circ}C$
No more than 1.0 ml or less than 0.1 ml sample plated (sample
or dilution)S
Liquified agar, 10 ml or more; medium temperature between 44° to 46°C
Merced mealum stored no longer than 3 hours before use
Only plates with between 30 to 300 colonies counted; when 1 ml
of undiluted sample is plated, colony density may be less
than 30
Only two significant figures recorded and calculated as
standard plate count/ml



GENERAL LABORATORY PRACTICES (Continued)

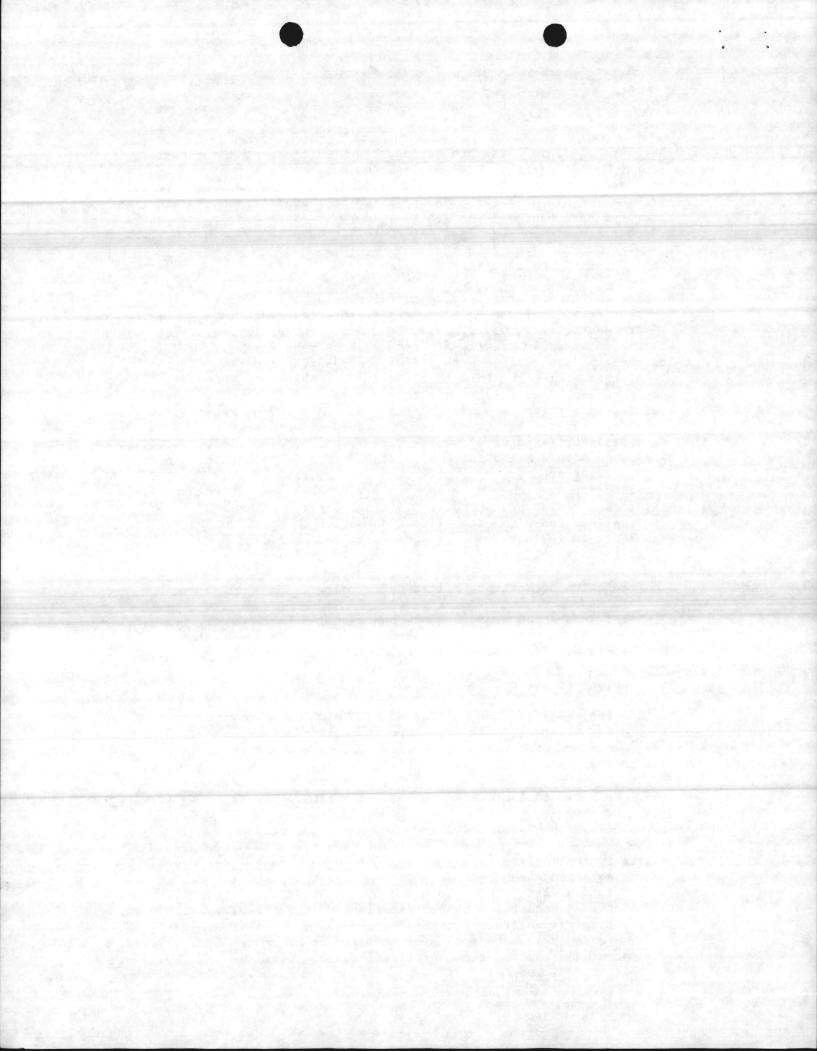
10.	Levine's Eosin Methylent Blue Agar (EMB)				
	Manufacturer <u>Difco</u>	Lot No. 701060 8/85			
	Medium composition 37.5g per liter <u>S</u> Final pH 7.1 <u>+</u> 0.2 <u>S</u>				
11.	Sterility Test Broth	<u>S</u>			
	Manufacturer Difco LTB	Lot No			

Will order T S B



METHODOLOGY

Metl M-Ei	ndo br	ogy specified in "Standard Methods" 14th edition, or EPA manual <u>S</u> oth, M-Endo agar, or Les Endo agar used in a single
In t	two-st sat	ep procedure <u>S</u> ep Les M-Endo procedure, MF incubated on lauryl tryptose broth curated absorbent pad for 1.5 to 2 hours at 35 \pm 0.5 C; then on Endo broth at Les Endo agar for 20 to 22 hours at 35 \pm 0.5 C <u>NA</u>
1.	Tot	al Coliform Membrane Filter Procedure
	San	ples containing excessive bacterial populations (greater than 200), confluency, or turbidity retested by the MPN procedure
	Abs San Tes	tration assembly sterile at start of each series
	MF	sterile buffered water <u>S</u> removed with sterile forceps, grasping outside effective
		filtering areaS rolled onto medium pad or agar so air bubbles are not trappedS tart and finish MF control test (rinse water, medium and supplies) run with each filtration series and results recordedS
	Whe	affected samples rejected and resampling requested
	a.	Incubation of Membrane Filter Cultures
		Total incubation time 22 to 24 hours at 35 ± 0.5 C
	b.	Membrane Filter Colony Counting
		Samples repeated when coliforms are "TNTC" or colony growth is confluent, possibly obscuring coliform development
		and/or detectionS Total coliform count calculated in density per 100 mlS Low power magnification device with fluorescent light positioned for maximum sheen visibilityS
	c.	Verification of Total Coliform Colonies
		All typical coliform (sheen) colonies or at least five randomly selected sheen colonies from each positive sample verified in lauryl tryptose broth and BGLBS Counts adjusted based on verificationS All atypical coliform (borderline sheen) colonies or at least randomly-selected colonies verified in LTB and BGLBS Counts adjusted based on verificationS
		Sheen colonies in mixed confluent growth reported and verified (optional)



METHODOLOGY (Continued)

d. MF Field Equipment

Manufacturer_	NA		Mo	del
Only standard	laboratory MF	procedures ad	lapted to	field
annlicati	OR	11	and the state of the state of the	

2. Total Coliform Most Probable Number Procedure

a. Presumptive Test

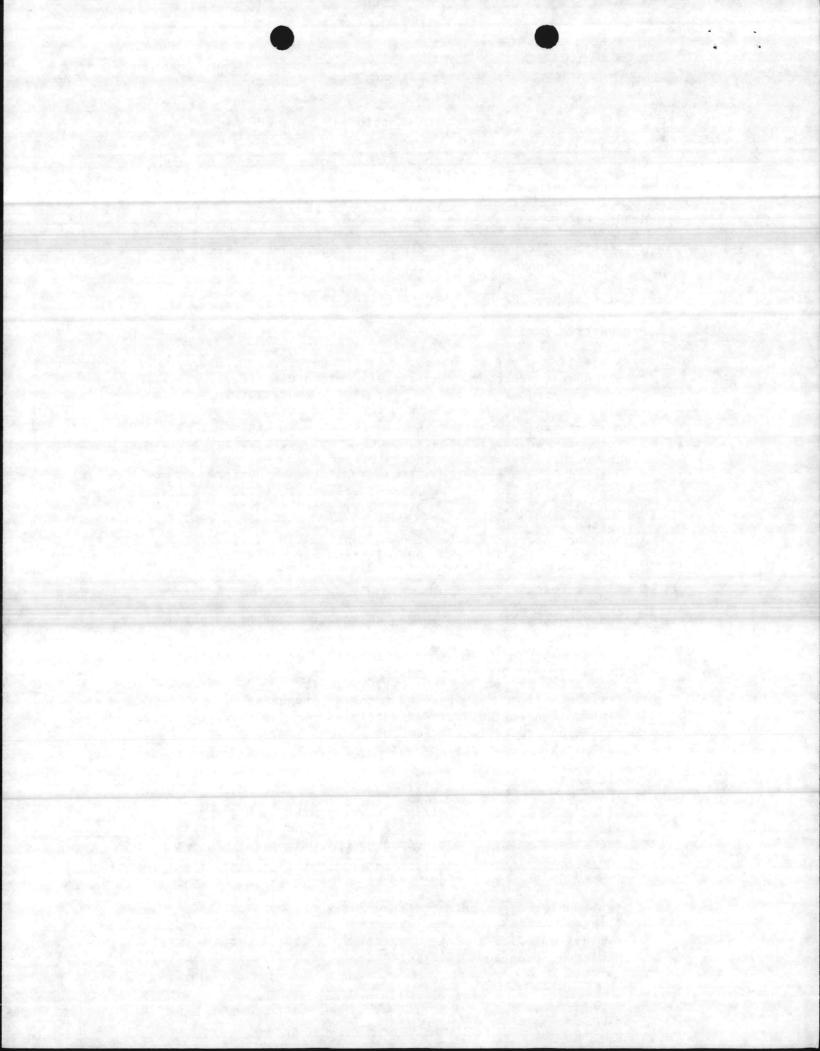
Five standard portions, either 10 or 100 mlS
Sample shaken vigorously immediately before testS
Tubes incubated at 35° + 0.5°C for 24 + 2 hoursS
Examined for gas (any gas bubble indicates positive test) S
Tubes that are gas-positive within 24 hours submitted
promptly to confirm testS
Negative tubes returned to incubator and examined for gas
within 48 + 3 hours; positives submitted to confirm
testS_
Public water supply samples with heavy growth and no gas
production confirmed for presence of supressed coliforms S
Adjusted count reported based upon confirmationS
Adequate test labeling and tube dilution coding (optional)

b. Confirmed Test

Presumptive positive tube gently shaken or mixed by rotating S
One loopful or one dip of applicator transferred from
presumptive tube to BGLB S
Incubated at $35^{\circ} + 0.5^{\circ}C$: checked within 24 hours + 2
hours for gas productionS
Positive confirmed tube results recorded; negative
tubes reincubated and read within 48 + 3 hours
Confirmation procedure carried out every 3 months on one
sample from each problem water supplyS

c. Completed Test

Applied to 10 percent of all positive samples each quarter
Applied to all positive confirmed tubes in each test completed S
Positive confirmed tubes streaked on EMB plates for
colony isolationS
Plates adequately streaked to obtain discrete coloniesS
Incubated at 35° + 0.5°C for 24 + 2 hoursS
Typical nucleated colonies, with or without sheen on EMB
plates selected for completed test identification S
If typical colonies absent, atypical colonies selected
for completed test identificationS



METHODOLOGY (Continued)

c. Completed Test (Continued)

If no colonies or only colorless colonies appear, confirmed
 test for that particular tube considered negative......S.
An isolated typical colony or two atypical colonies
 transferred to lauryl tryptose broth......S.
Incubated at 35^o + 0.5^oC; checked for gas within 48 + 3 hours....S.
Cultures producing gas in lauryl tryptose broth within
 48 + 3 hours are considered coliforms......S

3. Analytical Quality Control

a.	A	record	d of	analytical	quality	control	tests	available	
		for	revi	lew					x

Duplicate analyses

Duplicate analyses run on positive polluted samples not to exceed 10 percent but a minimum of one per month (optional)

Positive Control Samples

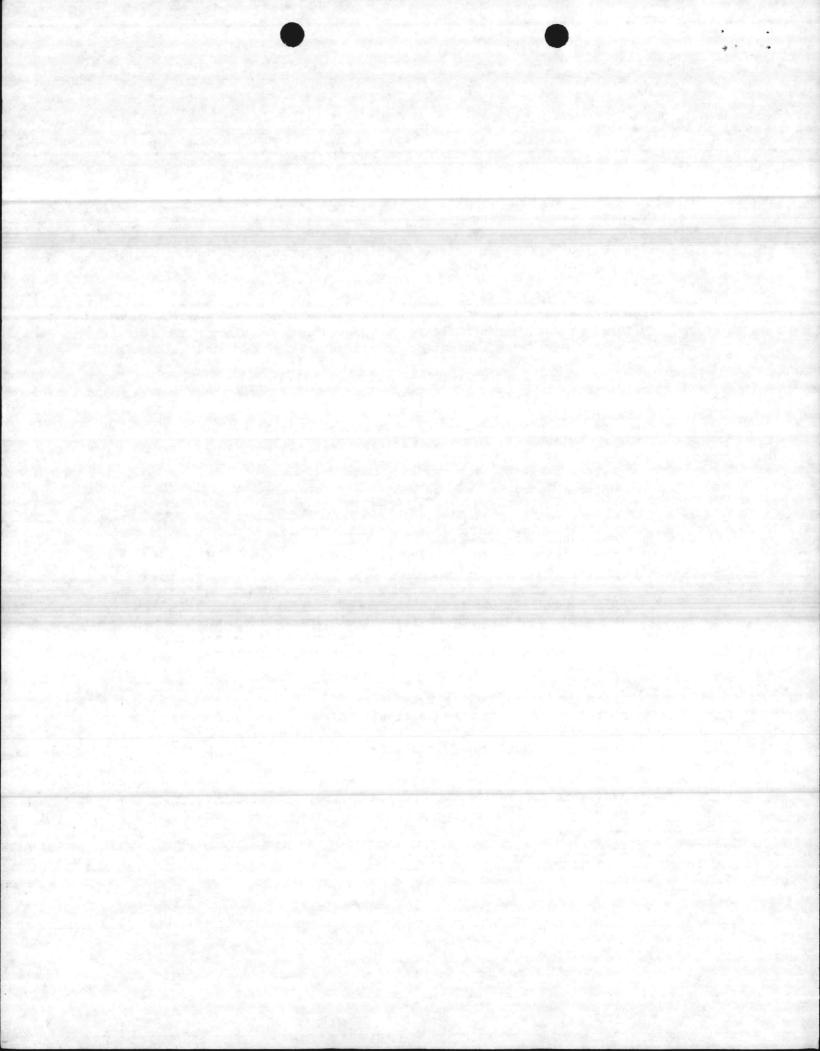
One positive control sample (polluted water) run each month (optional)

Colony Counting (If more than one Analyst in Laboratory) Two or more analysts count sheen colonies; all colonies are verified analysts' counts compared to verified counts; procedure is carried out at least once per month. (optional)

Check Analyses by State Laboratories A minimum of samples proportional to the local laboratory work

load processed by State Laboratory (see criteria for recommendations) (optional)

b. An outline of the quality control efforts of the laboratory available for review......S



SAMPLE COLLECTION, HANDLING, AND PRESERVATION

Representative samples of potable water distribution system......<u>S</u> Minimal sampling frequency as specified in the National Interim Primary Drinking Water Regulations.....<u>S</u> Sample collector trained and approved as required by State regulatory authority or its delegated representative.....<u>S</u>

1. Sample Bottles

Sodium thiosulfate, (10 mg per 100 ml of sample) added to
sample bottles before sterilizationS
Ample air space remains after sample collected to allow for
adequate mixingS

2. Sampling

Sample collected after maintaining a steady flow for	2 to 3 min
to clear service line	S
Tap free of aerator, strainer, hose attachment, water	
purification, or other devices	S
Samples refrigerated when possible during transit and	
periods in the laboratory (optional)	

3. Sample Identification

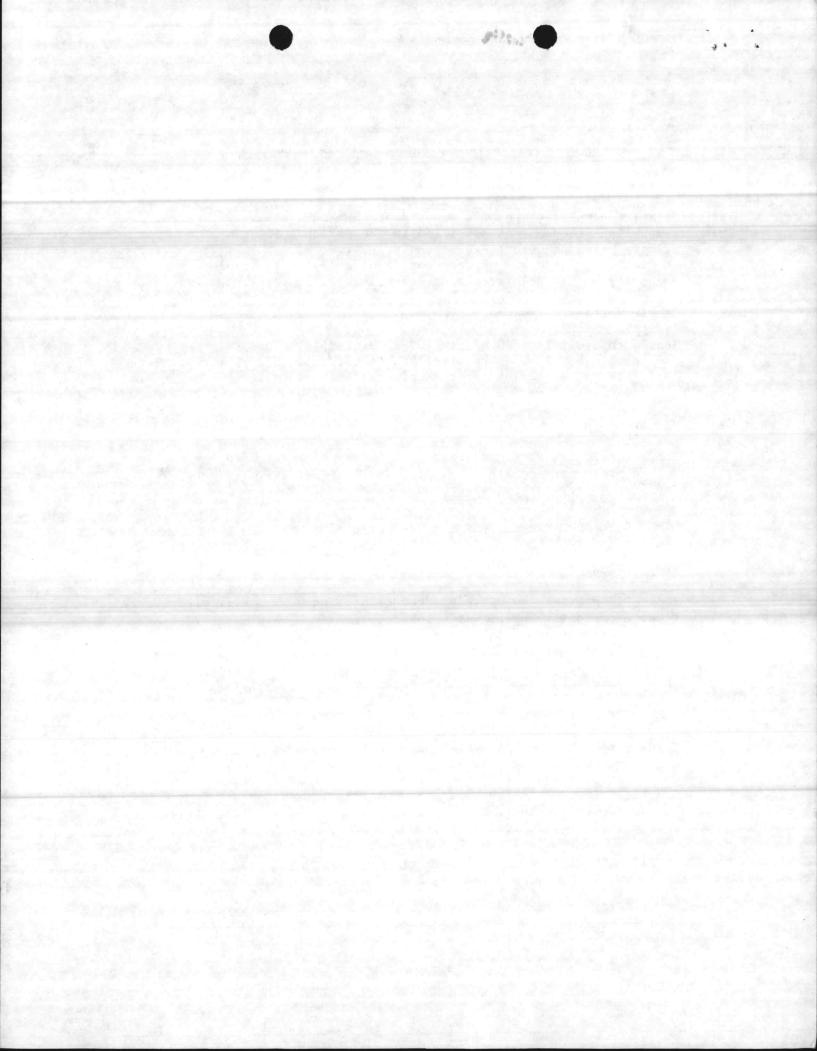
Sample identified immediately after collectionS
Identification includes, water source, location, water supply
identification number, time and date of collection, and
collector's name; insufficiently identified samples
discardedS
Chlorine residual where applicableS

4. Sample Transit Time

Transit time for potable water samples sent by mail or
commercial transportation, not in excess of 30 hoursNA
No sample processed after 48-hour transit/storageS
Samples delivered to laboratory by collectors examined the
day of collectionS
Data marked as questionable on samples analyzed after 30 hoursS

5. Sample Receipt in Laboratory

Sample logged in when received in laboratory, including date and time of arrival and analysis.....S Chain-of-custody procedures required by State regulations followed....S



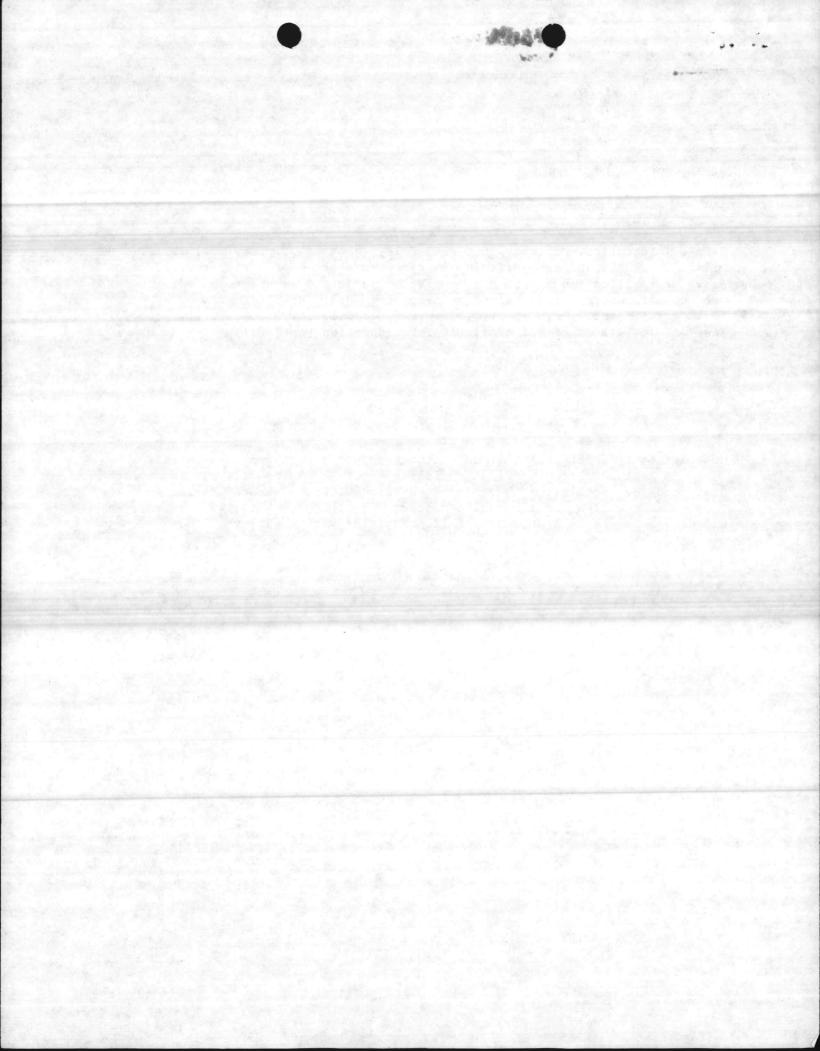
DATA REPORTING

Sample information and laboratory data fully recorded......S Direct MF counts and/or confirmed MPN results reported promptly......S After MF verification and/or MPN completion, adjusted counts reported.....S One copy of report form retained in laboratory or by State program for 3 years.....S

Test results assembled and available for inspection (optional)

ACTION RESPONSE TO LABORATORY RESULTS

Unsatisfactory test results given action response and resampled as defined	
in National Interim Primary Drinking Water Regulations S	5
State and responsible local authority notified within 48 hours after	
check samples confirm coliform occurrence	5
All data reported to State and local authorities within 40 days	S



OPNAV 5216/144A (Rev. 8-81) \$/N 0107-LF-052-2320



DATE: 29 August 1983

FROM: ElSupervisory Chemist, Quality Control Lab, Environmental Branch

TO: Supervisory Ecologist, Environmental Branch

SUBJ: State Inspection of Microbiology Laboratery

1. Don Beesley, Certification Inspector from the State, called today. He siddithat he was comming for an inspection on 8 September 1983. He said he will arrive around 1300. I Told him to report in at Bldg 1103 and to ask for you.

Elizabeth A. Betz Supervisory Chemist

